

**Arkansas Water Resources Center  
Annual Technical Report  
FY 2010**

# Introduction

The Arkansas Water Resources Center located at the University of Arkansas, Fayetteville, Arkansas is part of the network of 54 water institutes established by the Water Resources Research Act of 1964. Since its formation, the Arkansas Water Resources Center (AWRC) program in cooperation with the US Geological Survey and the National Institute for Water Resources has focused on helping local, state and federal agencies understand, manage and protect water resources within Arkansas. AWRC has contributed substantially to the understanding and management of water resources through scientific research and training of students. Center projects have focused on topics concerned with water quality of surface water and ground water, especially non-point source pollution and sensitive ecosystems. AWRC helps organize research to insure good water quality for Arkansas today and in the future.

The AWRC focuses its research on providing local, state and federal agencies with scientific data and information necessary to understand, manage and protect water resources within Arkansas. AWRC cooperates closely with colleges, universities and other organizations in Arkansas to address the state's water and land-related issues, promote the dissemination and application of research results, and provide for the training of scientists in water resources. Each year, several research faculty participate in AWRC projects with the help of students who gain valuable experience doing environmentally related work across the state. AWRC research projects have studied irrigation and runoff, innovative domestic wastewater disposal systems, ground water modeling and landuse mapping, erosion and pollution, water quality and ecosystem functions.

The Center provides support to the State's water research by acting as a liaison between funding groups and the scientists, and then coordinates and administers grants once they are funded. Accounting, reporting and water analyses are major areas of support offered to principal investigators. The AWRC has historically archived reports of water resource studies funded by the 104B program or through the Center on its website.

In addition, the AWRC sponsors an annual water conference held in Fayetteville, Arkansas each spring, drawing over 100 researchers, students, agency personnel and interested citizens to hear about results of current research and hot topics in water resources throughout the state. AWRC also co-sponsors short courses and other water-related conferences in the state and region. In addition, AWRC maintains a technical library containing over 900 titles, many of which are online. This valuable resource is utilized by a variety of user groups including researchers, regulators, planners, lawyers and citizens.

The AWRC also maintains a modern water quality laboratory that provides water analyses for researchers, municipal facilities, and watershed stakeholders; farmers and other citizens submit samples through the cooperative extension service. This laboratory is certified through the Arkansas Department of Environmental Quality for the analysis of surface and ground water samples.

The AWRC has a technical advisory committee made up of professionals from educational institutions, environmental organization, water supply districts, and government agencies throughout Arkansas. This committee has the opportunity to evaluate proposals submitted annually to the USGS 104B program, to recommend session topics included in the annual research conference, and to provide general advice to the AWRC Director and staff.

## Research Program Introduction

Each year, several researchers participate in 104B projects funded through the Arkansas Water Resources Center (AWRC), and these projects are completed with the help of students in water and environmentally related fields. The research projects funded through the AWRC have studied a broad range of environmental and water issues facing Arkansas, including irrigation and rainfall–runoff, innovated domestic wastewater disposal, groundwater modeling and land use mapping, erosion and nonpoint source pollution, water quality and ecosystem function. The AWRC has given priority to solid scientific research proposals submitted by faculty to the 104B program; the intent has been to provide seed data to researchers such that larger proposals can be developed and submitted to extramural funding sources. The AWRC has funded several projects using 104B funding that have resulted in the award of extramural grants to continue the base research.

To formulate a research program relevant to state water issues, the Center works closely with state and federal agencies, and academic institutions. An advisory committee, composed of representatives from state and federal agencies, industry and academia, provides guidance for the Center. The technical advisory committee plays an important role in insuring that the water institute program (section 104) funds address current and regional issues. The priority research areas of the AWRC base program directly related to the program objectives of the Water Resources Research Act, including research that fosters improvements in water supply, explores new water quality issues, and expands the understanding of water resource and water related phenomena.

# Determination of the magnitude of mercury methylation in the water column of a high organic carbon river, lower Ouachita River, Union and Ashley Counties, Arkansas

## Basic Information

<b>Title:</b>	Determination of the magnitude of mercury methylation in the water column of a high organic carbon river, lower Ouachita River, Union and Ashley Counties, Arkansas
<b>Project Number:</b>	2010AR248B
<b>Start Date:</b>	3/1/2010
<b>End Date:</b>	2/28/2011
<b>Funding Source:</b>	104B
<b>Congressional District:</b>	3
<b>Research Category:</b>	Water Quality
<b>Focus Category:</b>	Toxic Substances, Surface Water, Hydrogeochemistry
<b>Descriptors:</b>	None
<b>Principal Investigators:</b>	Phil D Hays, Stephen K. Boss, John Van Brahana, Ralph K. K Davis

## Publications

1. Schenk, Liam. 2011. Geochemical Controls on Mercury Methylation in Backwaters of a Gulf Coastal Plain River System, Implications for Water Column Processes. Thesis, Master of Science in Geology, University of Arkansas, Fayetteville, Arkansas.
2. DETERMINATION OF THE MAGNITUDE OF MERCURY METHYLATION IN THE WATER COLUMN OF A RIVER WITH HIGH DISSOLVED ORGANIC CARBON, LOWER OUACHITA RIVER, UNION AND ASHLEY COUNTIES, ARKANSAS: SCHENK, Liam N., Department of Geosciences, University of Arkansas, 113 Ozark Hall, Fayetteville, AR 72701, lschenk@uark.edu and HAYS, Phillip, U.S. Geological Survey, Ozark Hall 113, Fayetteville, AR 72701

**Project Title:** Determination of the Magnitude of Mercury Methylation in the Water Column of a High Organic Carbon River, Lower Ouachita River, Union and Ashley Counties, Arkansas

**Project Team:** Dr. Phil D. Hays, Arkansas US Geological Survey

Dr. Stephen K. Boss, Geosciences, University of Arkansas

Dr. John Van Brahana, Geosciences, University of Arkansas

Dr. Ralph K. Davis, Geosciences, University of Arkansas

**Interpretative Summary:** The abundance and distribution of mercury and methyl mercury were investigated at three sites in the lower Ouachita River in the summer of 2010 in an effort to provide the first characterization of the extent of mercury contamination in this river system, and to investigate the potential for mercury methylation in the water column of backwaters off of the main channel. Results showed that filtered methyl mercury was positively correlated to dissolved organic carbon ( $r_2=0.76$ ) for water samples taken from the bottom 1 ft of the water column at three sites, suggesting the importance of dissolved organic carbon in mercury methylation. Concentrations of filtered methyl mercury and filtered total mercury in the bottom-water were significantly different ( $P=0.039$  and  $P=0.022$  respectively) at two of the sample sites located approximately 14 river miles apart. Sulfide concentrations of 74.0-142.7 micrograms/liter indicate sulfate reduction was occurring in the bottom water or at the sediment-water interface, yet filtered and particulate methyl mercury concentrations were not significantly correlated to sulfide concentrations. The occurrence of sulfides in the bottom-water is important as sulfate-reducing bacteria are most commonly associated with mercury methylation. Water chemistry results for one site including total iron (39.8 milligrams/liter), high dissolved organic carbon (13.52 milligrams/liter), the highest filtered methyl mercury concentration observed for the study (1.90 nanograms/liter), and no detectable sulfate suggests the predominance of iron reduction at this site. Microbial iron reduction is also a known mercury methylation pathway. Total mercury concentrations for two of seven samples exceeded the Arkansas numeric water quality standard for total recoverable mercury in water (12 nanograms/liter), at concentrations of 13.76 and 13.99 nanograms/liter. These data provide evidence that availability of dissolved organic carbon affects mercury methylation at all three of the sites, and that iron reduction may contribute to mercury methylation at one of the sites. No correlation between sulfide and dissolved methyl mercury was observed, suggesting sulfate reduction may not be the driving process for mercury methylation at all our study sites, and indicating the presence of multiple controls on mercury methylation in this river system.

**Introduction:** Increased scientific knowledge on mercury (Hg) sources, transport, deposition and cycling, and the toxic effects of Hg species on human populations has led to growing concern over Hg contamination of aquatic systems in recent years. Hg is naturally present in the environment, but human activities such as the combustion of fossil fuels for power generation have increased the amount of Hg cycling through land, atmosphere, and ocean systems (N. E. Selin 2009). As such, atmospheric deposition of Hg is increasing in marine systems (Sunderland, et al. 2009), as well as in riverine systems (DeLongchamp, et al. 2009). Riverine systems in some regions such as those draining cinnabar mining districts are at even greater risk of contamination due to exposure and mobilization of Hg during cinnabar mining (Holloway, et al. 2009).

Of primary concern is the formation of the most toxic form of Hg, methylmercury (MeHg). In aquatic systems, Hg can be deposited by either wet or dry deposition in its elemental (Hgo) and ionic (Hg(II)) forms. Hgo and Hg(II) can then be transformed into toxic and highly bioavailable MeHg, the result of processes largely carried out by anaerobic bacteria (Marvin-Dipasquale, et al. 2009). Epidemiological studies have linked exposure to MeHg in pregnant women to neurological and developmental effects in their offspring (Mergler, et al. 2007), (Clarkson 1990). High degrees of human exposure to MeHg most commonly results from the consumption of high trophic-level predaceous fish such as tuna and swordfish in marine systems, and black bass and piranha in freshwater systems.

The objective of this project is two-fold. The first objective is to provide the first detailed characterization of the occurrence and extent of Hg contamination in backwaters of the lower Ouachita River system, and second, to examine the geochemical controls on MeHg in the water column. A detailed multimedia, multi chemical-species sampling scheme was implemented to characterize the relation between organic carbon and Hg methylation. Field parameters along with sulfide, sulfate, and iron concentrations were assessed in the bottom 1-ft of the water column (hereafter referred to as bottom water) to provide insight into oxidation-reduction (redox) conditions that dominate the system and provide evidence for the presence of anaerobic bacteria known to be responsible for Hg methylation. Determination of the controls on MeHg occurrence in the bottom water is achieved by comparing total Hg (THg) and MeHg concentrations to field parameters, DOC, sulfide, iron, sulfate and MeHg sediment concentrations. The characterization of Hg contamination at three backwater sites on the lower Ouachita River provides detailed, state-of-the-science Hg data that give insight into the extent of Hg contamination.

**Methods:** Three sites (RL-2, OR-2 and OR-11) were chosen as the most likely candidates for MeHg production in the Ouachita River System. Sampling of all three sites was conducted from late July to early August 2010. This time was selected to target the season that would have the highest ambient temperatures and water temperatures of the year, and lowest water flow, and thus the most likely time for stratification to occur in the water column. Surface water samples were collected three times at RL-2 and OR-2 and twice at OR-11 and analyzed for THg, MeHg, DOC, and physico-chemical parameters; sediment samples were collected and analyzed for MeHg. A diurnal sampling event was implemented at Or-2 to determine any potential fluctuations of Hg and sulfides over a 24-hour period.

Non-parametric statistics were used to compare median values of bottom-water concentrations of HG species and other parameters between sites. Sign-rank and rank-sum tests were used to test the equality of median values, and one-way ANOVA's were used to test for equality of means. Statistical significance was set at  $\alpha=0.05$ .

**Results:** All three sites exhibited stratification with respect to temperature and DO, with high temperature and high DO in the shallow depths, and lower temperatures and anoxic conditions in the bottom-water. pH values did not vary much between sites or between water surface and bottom water, while specific conductance for all three sites increased with increasing depth in the water column.

Concentrations of Hg compiled from all three sites show increases from the dissolved MeHg (FMeHg) fraction to total Hg (THg), with the lowest FMeHg concentrations occurring at 0.05 ng/L, just above the MDL (0.04 ng/L), and the highest Hg concentrations occurring in the THg fraction at 13.99 ng/L, calculated as the sum of filtered and particulate THg.

Mercury concentrations in all size fractions and speciation varied between sites (Figure 1). RL-2 had the highest dissolved MeHg and THg as compared to the other sites. OR-11

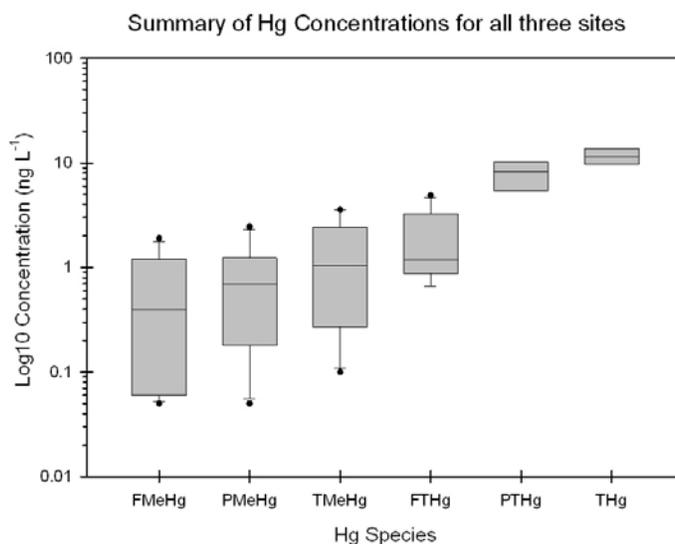


Figure 1: Hg concentrations combined from all three site including mid point water column and bottom-water samples. n=11 for FMeHg, PMeHg, TMeHg, FTHg, n=7 for PTHg, THg

had the lowest FMeHg, PMeHg, and FTHg, but had the highest PTHg and THg. Hg concentrations at OR-2 were between the other two sites for all size fractions and species. FMeHg and PMeHg at OR-2 showed variability in concentrations between sample days shown by the error bars in Figure 2 representing standard deviation of samples collected on three separate sample days.

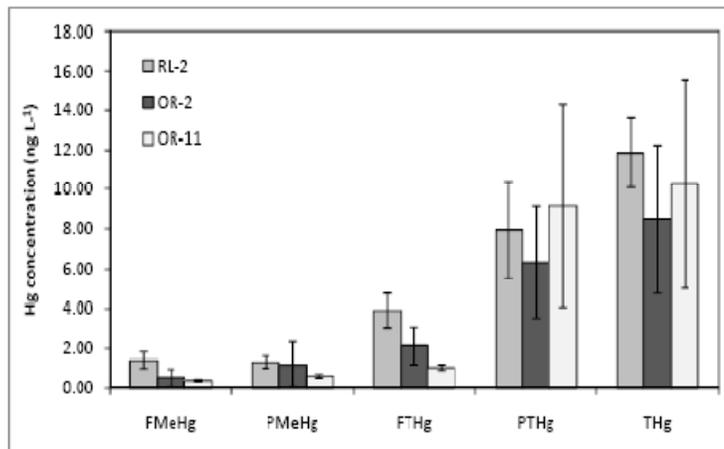


Figure 2: Bottom water concentrations of Hg size fractions and species at all three study sites. N=3 for all sites. Error bars represent standard deviations of samples collected on separate days.

Non-parametric statistics were used to compare bottom-water samples of Hg species and size fractions between sites. There was no statistical difference of median values at the bottom-water for any of the Hg species and size fractions when using rank-sum or sign-rank statistical tests. One-way ANOVA's used to test the difference of means resulted in statistical differences of FMeHg in the bottom-water between RL-2 and OR-11 ( $P=0.039$ ) and FTHg between RL-2 and OR-11 ( $P=0.022$ ) only. All other species and size fractions were not statistically different at the bottom-water between sites. MeHg and THg were positively correlated at the bottom-water for filtered samples, but not significantly correlated for total MeHg (TMeHg) and THg.

Of the three study sites, RL-2 exhibited the highest dissolved MeHg and THg, and particulate MeHg concentrations in the bottom water (Figure 2). Additionally, the highest DOC concentrations were encountered at this site (mean 13.37 mg/L,  $n=3$ ), and the highest total iron concentration (39.8 mg/L). These data provide evidence for geochemical controls on Hg methylation at this site, as high DOC is often related to high rates of Hg methylation, and high iron concentration gives evidence for redox processes that may have been controlling methylation.

The highest dissolved MeHg concentrations of all three study sites occurred at RL-2 (mean 1.43 ng/L,  $n=3$ ), which also had the highest DOC concentrations (mean 13.37 mg/L,  $n=3$ ), highlighting the importance of DOC in Hg methylation processes at this site. The positive linear correlation with DOC and dissolved MeHg (Figure 34) indicates that DOC may not be inhibiting Hg methylation at this site, and is

providing an energy source to methylating bacteria, potentially iron reducers, owing to the high iron concentration encountered at this site.

THg and MeHg concentrations at all three study sites on the lower Ouachita River are typically within the range of Hg values reported in the literature for the southeastern United States, and median and mean values exceeded reported concentrations in many areas. The highest dissolved MeHg concentration on the lower Ouachita River system was 1.90 ng/L at RL-2, highlighting the high rates of MeHg production. Although the three study sites on the lower Ouachita are not technically classified as wetlands, they can experience similar fluctuations in water levels as they are hydraulically connected to the main channel of the river, which fluctuates with seasons. These fluctuations do not expose large areas of sediment as compared to most freshwater wetlands. The concentrations of dissolved MeHg at the lower Ouachita sites are much higher than many/most of the regional concentrations, suggesting high rates of methylation, potentially due to similar conditions as explained by Hall, et al. (2008).

**Conclusions:** Data generated from this study show the spatial variability in geochemistry at the study sites, which has a direct effect on MeHg production. Sites with high DOC had higher concentrations of dissolved MeHg as evidenced by positive correlation between these two constituents. Measureable sulfides in the bottom water at all three sites give evidence for sulfate reduction, yet high absolute values of redox potential indicate that redox potential is not low enough in the bottom-water at the study sites to allow sulfate and iron reduction.

The observed positive linear relation between dissolved MeHg and DOC at the bottom water at all three sites indicates an important influence of DOC on Hg methylation. This relation can be explained by DOC acting as an energy source that stimulates microbial activity, and by low pH in the bottom water providing protons to compete for negatively charged binding sites in DOC that would otherwise be utilized by Hg, thus leaving Hg bioavailable for methylation (Barkay, Gillman and Turner 1997). Channel morphology also plays an important role in the distribution of DOC, and in site specific stratification characteristics.

Measurable sulfide detected at the bottom water at all three sites suggest that sulfate reduction and associated MeHg production may be occurring either in the anoxic water at the base of the water column, at the sediment-water interface, or in sediment pore water with sulfide and MeHg moving out of sediment into the overlying water column. However, ORP values do not show the potential for sulfate reduction in the anoxic bottom waters at the lower Ouachita River sites. Given the conflicting evidence

of absolute ORP, further data are needed at corroborate the occurrence of sulfate reduction in the bottom water at these sites including a larger sulfide data set, dissolved iron analysis, and a larger sulfate data set.

Of the three study sites, RL-2 exhibited the highest filtered MeHg, THg, and particulate MeHg concentrations in the bottom water as well as the highest DOC concentrations (mean 13.37 mg/L, n=3). Measureable sulfide at this site indicates that sulfate reduction may be occurring concurrently with iron reduction. It is therefore possible that multiple microbial communities that methylate Hg are responsible for the high concentrations of MeHg at RL-2, potentially even FeRB.

Assessment of seasonal fluctuations of Hg from existing USGS data at sites proximal to the lower Ouachita River sampling sites show peaks in the occurrence of filtered THg and MeHg during late spring, indicating seasonal controls on MeHg production. An increase in the supply of DOC during high precipitation or flood events may increase MeHg production by enhancing microbial activity.

This study provides crucial data describing the extent of Hg contamination in Arkansas, with two of eight bottom-water samples exceeding the numeric water quality standard of 12 ng/L total recoverable Hg in water. As atmospheric Hg deposition increases across the country, the Hg issue in Arkansas only stands to become more prominent, giving impetus for additional research to be conducted on this important environmental issue.

#### **References:**

Barkay, Tamar, Mark Gillman, and Ralph Turner. "Effects of Dissolved Organic Carbon and Salinity on Bioavailability of Mercury." *Applied and Environmental Microbiology* 63, no. 11 (1997): 4267-4271.

Hall, B.D., G.R. Aiken, D.P. Krabbenhoft, M. Marvin-DiPasquale, and C.M. Swarzenski. "Wetlands as principal zones of methylmercury production in southern Louisiana and the Gulf of Mexico region." *Environmental Pollution* 154 (2008): 124-134.

**Thesis or Dissertation:** Schenk, Liam. 2011. Geochemical Controls on Mercury Methylation in Backwaters of a Gulf Coastal Plain River System, Implications for Water Column Processes. Thesis, Master of Science in Geology, University of Arkansas, Fayetteville, Arkansas.

# Assessment of water quality and stream bank stability following BMP implementation on the upper Strawberry River watershed

## Basic Information

<b>Title:</b>	Assessment of water quality and stream bank stability following BMP implementation on the upper Strawberry River watershed
<b>Project Number:</b>	2010AR249B
<b>Start Date:</b>	3/1/2010
<b>End Date:</b>	2/28/2011
<b>Funding Source:</b>	104B
<b>Congressional District:</b>	AR-001
<b>Research Category:</b>	Not Applicable
<b>Focus Category:</b>	None, None, None
<b>Descriptors:</b>	None
<b>Principal Investigators:</b>	Jennifer L Bouldin

## Publication

1. Brueggen, T.R. In progress. Effects of Best Management Practices on the Upper Strawberry River Watershed, Fulton CO, AR. Environmental Sciences, Arkansas State University.

**Project Title:** Investigation of Land Use and Best Management Practices on the Strawberry River Watershed

**Project Team:** Dr. Jennifer Bouldin, Environmental Biology, Arkansas State University  
Teresa Brueggen, Environmental Sciences, Arkansas State University

**Interpretative Summary:** Best Management Practices (BMPs) including exclusion of cattle from waterways, providing alternative watering facilities, and use of no-till planting methods have been put into place on three creeks in the upper watershed of the Strawberry River, AR. This study incorporates physical, biological and chemical analyses to determine the effects of the implemented BMPs on water and sediment quality of the three creeks. Protection of upper headwater streams will improve ecosystem integrity downstream in this Ecologically Sensitive Waterbody. This study has the potential to expand the knowledge base of improved water quality from stream-side agricultural BMPs.

**Introduction:** The Strawberry River Watershed is located in the Ozark Highland Ecoregion of Arkansas and defined as an Extraordinary Resource Water, Ecologically Sensitive Water Body, and Scenic Waterway (ADEQ, 2008). The waters of the Strawberry River support a diversity of species including the endogenous Strawberry River Darter, diverse communities of aquatic macroinvertebrates including several ranked or listed freshwater mussels (Harp and Robinson, 2006). The ADEQ (2008) defines the designated uses for the Strawberry River as Primary and Secondary Contact Recreation, Domestic, Industrial and Agricultural Water Supply. Seven reaches of the Strawberry River Watershed are included in the 303d list as not supporting aquatic life due to excess turbidity (ADEQ, 2008). Land use in the watershed is primarily forested (56.5%) and grassland (35.3%) (ADEQ, 2003), with livestock grazing and hay harvesting for livestock common among the grassland owners. Grazing practices often do not include fencing from the streambed leading to increased bank sloughing. Best management practice implementation is presently underway in the upper watershed and landowner participation is encouraged through an EPA 319 grant issued to the Fulton County Conservation District and Arkansas State University. Upstream and downstream monitoring sites are located on Little Strawberry, Greasy Creek and Sandy Creek.

**Methods:** Erosion pins were used to assess bank stability and estimate sediment transport from bank erosion. Multiple pins were installed perpendicular into the stream bank. These pins will be installed at

the active bank and above the active bank determined at each designated sampling location (Zaimes et al., 2005). A survey of each stream reach quantified the extent of stream with bank instability.

Benthic surveys will be performed with D-frame nets using the traveling kick method. Organisms will be keyed to species according to Merritt et al. (2008) whenever possible and 10% of samples will be referred to a benthic taxonomist for Quality Assurance of identification.

Enumeration of *E. coli* and measures of chlorophyll *a* will be determined monthly. *Escherichia coli* and chlorophyll *a* concentrations will be determined using the filtration technique in accordance with the American Public Health Association (APHA, 2005).

Aqueous and sediment toxicity studies will be performed in the fall and spring. Bioassays will be used to measure the presence of toxicity. *Ceriodaphnia dubia* and *Pimphales promelas* will be used in whole effluent toxicity (WET) 7-d chronic tests, in accordance with the EPA guidelines (2002) to determine aqueous toxicity. Presence of sediment toxicity will be measured using *Chironomus dilutus* with a 10-d acute toxicity test in accordance with EPA guidelines (2000).

**Results:** Approximately 5480 m of stream bank was assessed within the Little Strawberry Creek. It was determined that there were 24 sites of severe or very severe erosion totaling approximately 746 m of stream bank. Approximately 6340 m of stream bank were assessed of Greasy Creek. In this stretch 16 sites were determined as severely or very severely eroded totaling approximately 500 m. Approximately 13260 m of stream bank was assessed of Sandy Creek. Twenty two sites were classified with severe or very severe erosion totaling approximately 505 m. An assessment of the erosion pins was performed in October 2010.

In spring and fall 2010 benthic macroinvertebrate collections, 2705 and 1328 total organisms were collected, respectively (Fig 1). Sandy Creek upper site was not sampled fall 2010 due to dry conditions. This included the following: Coleoptera, Diptera, Ephemeroptera, Hemiptera, Megaloptera, Odonata, Plecoptera, Trichoptera, Decapoda, and Mollusca. Total family diversity between sites for spring and fall ranged from 2-24 and 6-19, respectively (Fig 2).

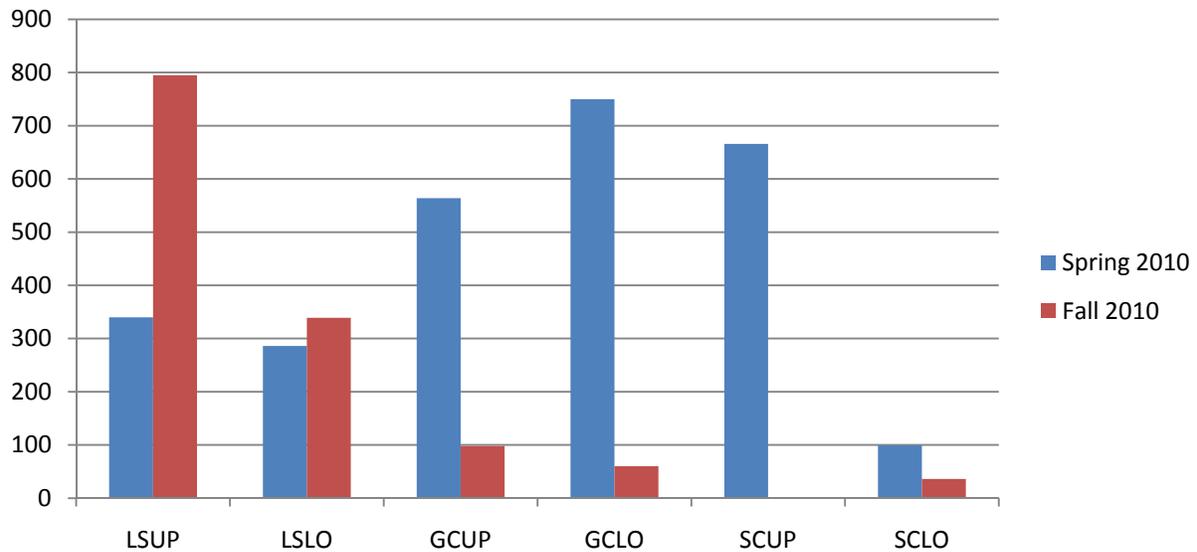


Figure 1. Total number of organisms collected at monitoring sites in the upper watershed of the Strawberry River. Little Strawberry (LS), Greasy Creek (GC), Sandy Creel (SC), upper location (UP) and lower location (LO). SCUP not sampled fall 2010 due to dry sampling location.

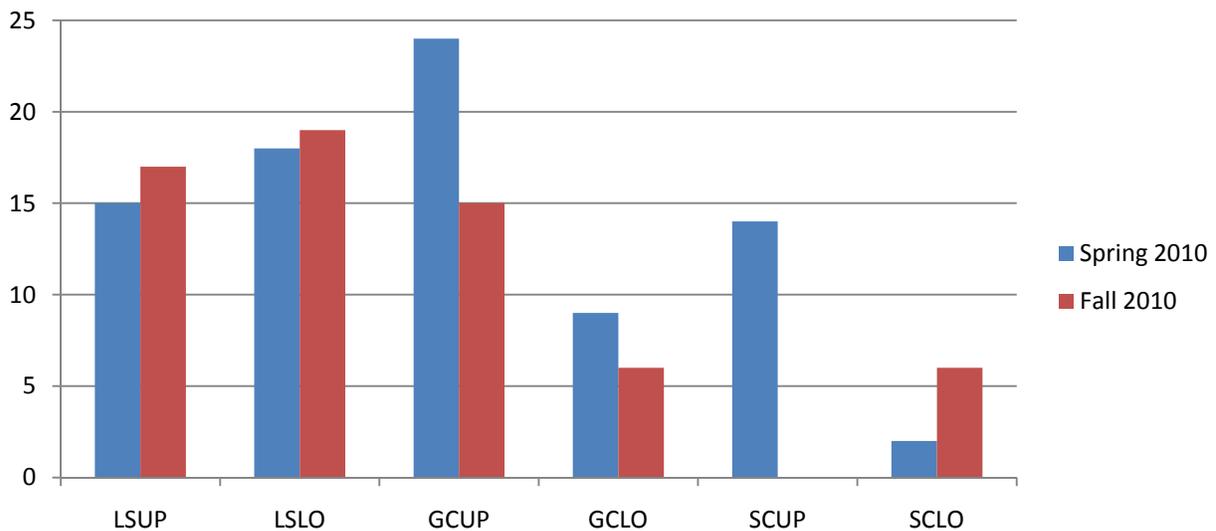


Figure 2. Total number of families identified at monitoring sites in the upper watershed of the Strawberry River. Little Strawberry (LS), Greasy Creek (GC), Sandy Creel (SC), upper location (UP) and lower location (LO). SCUP not sampled fall 2010 due to dry sampling location.

*E. coli* mean values ranged from 51-215 colony forming units (CFUs). No single sample concentrations exceeded allowable limits (APCEC, 2010). Little Strawberry upper site indicated significant lethal aqueous toxicity using *P. promelas* in Spring 2010 and lethal sediment toxicity in Fall 2010.

**Conclusions:** This is an ongoing study; therefore, final conclusions are limited. It is evident that multiple years of analysis assessing physical, chemical and biological parameters are vital to evaluate the impact of implemented BMPs. Much variability can occur from year to year as environmental parameters outside of the researcher's control fluctuate (e.g. rainfall, temperature).

**References:**

- Arkansas Department of Environmental Quality. 2008. Arkansas' 2008 303(d) List of impaired waterbodies. Published by Arkansas Department of Environmental Quality. 18pp.
- Arkansas Department of Environmental Quality. 2003. Physical, chemical and biological assessment of the Strawberry River Watershed. Published by Arkansas Department of Environmental Quality. QA-03-12-01. 282pp.
- American Public Health Association. 2005. Standard methods for the examination of water and wastewater. 21st ed. American Public Health Association, Washington D.C. 1325pp.
- Arkansas Pollution Control and Ecology Commission (APCEC). 2010. Regulation no. 2. Regulation establishing water quality standards for surface water of the state of Arkansas. Arkansas Department of Environmental Quality. 124pp.
- Harp, G.L. and H.W. Robison. 2006. Aquatic Macroinvertebrates of the Strawberry river system in north-central Arkansas. *Journal of the Arkansas Academy of Science* 60:46-61.
- United States Environmental Protection Agency. 2000. Methods for measuring the toxicity and bioaccumulation of sediment-associated contaminants with freshwater invertebrates. United States Environmental Protection Agency, National Center for Environmental Publications (NSCEP), Cincinnati, OH. EPA 600/R-99/064.
- United States Environmental Protection Agency. 2002. Short-term methods for estimating the chronic toxicity of effluents and receiving waters to freshwater organisms. 4th ed. EPA 600/4-91/002.
- Zaimes, G. N., Schultz, R.C., Isenhardt, T.M., Mickleson, S.K., Kovar, J.L., Russell, J.R. and Powers, W.P. 2005. Stream bank erosion under different riparian land-use practices in northeast Iowa: AFTA 2005 Conference Proceedings, 1-10.

**Thesis or Dissertation:**

Brueggen, T.R. In progress. Effects of Best Management Practices on the Upper Strawberry River Watershed, Fulton CO, AR. Environmental Sciences, Arkansas State University.

# Denitrification, Internal N Cycling, and N Retention in River Impoundment Reservoirs

## Basic Information

<b>Title:</b>	Denitrification, Internal N Cycling, and N Retention in River Impoundment Reservoirs
<b>Project Number:</b>	2010AR252B
<b>Start Date:</b>	3/1/2010
<b>End Date:</b>	2/28/2011
<b>Funding Source:</b>	104B
<b>Congressional District:</b>	3
<b>Research Category:</b>	Water Quality
<b>Focus Category:</b>	Geochemical Processes, Nutrients, Sediments
<b>Descriptors:</b>	None
<b>Principal Investigators:</b>	Jefferson Thad Scott

## Publication

1. Grantz, E. 2011. Denitrification efficiency in water impoundment reservoirs, Department of Crop, Soil, and Environmental Sciences, University of Arkansas.

**Project Title:** NITROGEN RETENTION AND DENITRIFICATION EFFICIENCY IN WATER IMPOUNDMENTS

**Project Team:** J. Thad Scott, Department of Crop, Soil, and Environmental Sciences, Univ. of Arkansas

**Interpretative Summary:** Reactive nitrogen loss from human-impacted landscapes may be degrading water quality in downstream coastal environments. This project was conducted to determine what role water impoundment reservoirs play in attenuating reactive N from surface waters by transformation to N<sub>2</sub> gas via denitrification. Over the last two years we sampled three reservoirs in Northwest Arkansas to determine their capacity to store and transform reactive N. We collected intact sediment cores to quantify denitrification rates, created N mass balances, and developed a new method for estimating whole-ecosystem N<sub>2</sub> flux data from thermally stratified reservoirs. Our data suggest that reservoirs are indeed important N sinks and that a substantial portion of stored N is eventually denitrified.

**Introduction:** Denitrification in water impoundment reservoirs may remove substantial quantities of reactive N from surface waters, but few comprehensive denitrification studies have been conducted on reservoirs (David et al. (2006)). More work is needed to quantify denitrification rates in reservoirs and the factors that cause rates to vary across space and time. In particular, these studies should address what percentage of N retention is caused by denitrification. Developing a thorough understanding of the factors that control denitrification in reservoirs will allow us to maximize reservoir N retention through proper reservoir management.

In this study we measured sediment denitrification rates, seasonal ecosystem-scale denitrification, and N storage (mass balance) in three water impoundment reservoirs in Northwest Arkansas. The objective of the study was to quantify denitrification rates in these reservoirs and estimate what portion of stored N in the reservoirs is ultimately denitrified.

**Methods:** We used intact sediment cores collected from epilimnetic sediments during spring and summer stratification periods, and all sediments during winter mixing, to estimate the rate of denitrification in sediments. Denitrification on intact cores was estimated using a mass balance on N<sub>2</sub> concentrations occurring in the inflow and outflow of flow-through core chambers. N<sub>2</sub> concentrations were derived from the N<sub>2</sub>/Ar ratio, measured using membrane inlet mass spectrometry.

Hypolimnetic denitrification was estimated by monitoring the N<sub>2</sub>/Ar ratio in the hypolimnion of the three study reservoirs through the period of summer stratification in 2010. Briefly, water samples from 4m, 6m, and 8m were collected weekly and preserved immediately with zinc chloride. N<sub>2</sub>/Ar ratio of these samples was measured using MIMS as described previously. The rate of N<sub>2</sub> accumulation was estimated by assuming that Ar concentrations were controlled only by temperature and that the slope of a statistically significant linear regression of N<sub>2</sub> concentration versus time represented the N<sub>2</sub> accumulation rate.

Nitrogen retention in reservoirs was calculated as the difference between riverine N inputs and outputs. Nitrogen inputs to reservoirs were estimated using the rating curve method (Shivers and Moglen 2008). Briefly, a relationship between stream stage, streamflow, and total N concentrations were derived for inflowing and outflowing streams to estimate N inputs from continuously monitored stream stage. Yields from gauged streams were applied to ungauged streams to estimate whole-system inputs.

### Results:

Epilimnetic Sediment Denitrification – Average net sediment N<sub>2</sub> flux rates for all three lakes are provided in Figure 1. All three lakes exhibit net denitrification (i.e. positive net N<sub>2</sub> flux) during winter. This condition coincides with measurable nitrate-nitrogen (NO<sub>3</sub>-N) in the lakes during this time. Spring and summer net N<sub>2</sub> flux was either negative (i.e. net N<sub>2</sub> fixation in May 2010) or zero (i.e. equal amounts of denitrification and N<sub>2</sub> fixation). Nitrate concentrations in the epilimnion of these lakes during summer is below detection levels and is probably the limiting factor that inhibits denitrification and cause net zero sediment N<sub>2</sub> flux from epilimnetic sediments during spring and summer stratification.

Hypolimnetic Denitrification – Hypolimnetic N<sub>2</sub> accumulation from one of the study lakes is shown in Figure 2. N<sub>2</sub> gas accumulated at a linear rate throughout

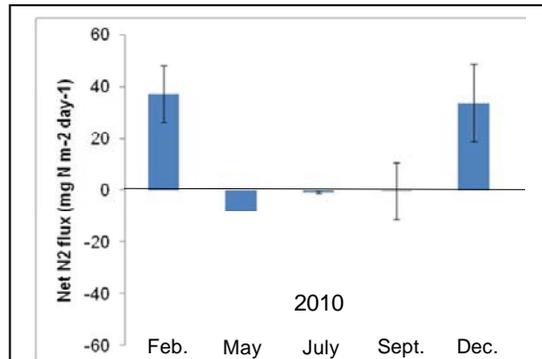


Figure 1. Average Net N<sub>2</sub> flux rates derived from intact core experiments on all three study lakes.

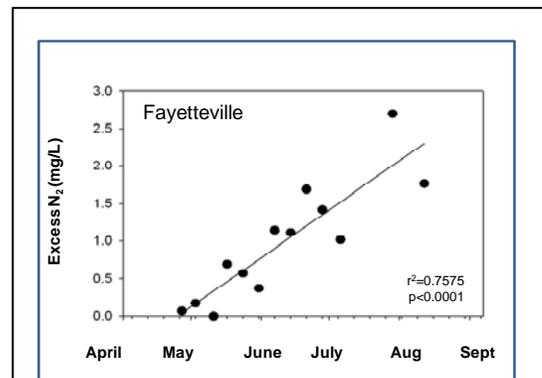
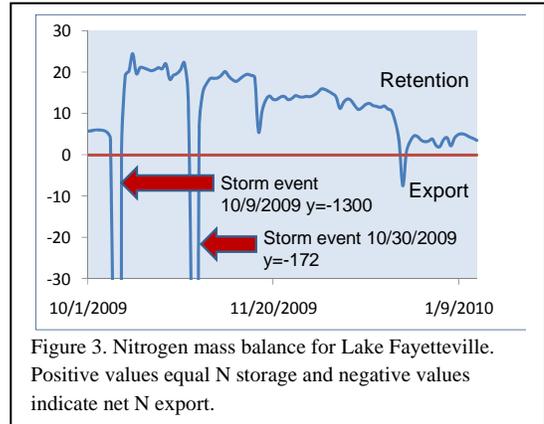


Figure 2. Hypolimnetic N<sub>2</sub> accumulation in Lake Fayetteville, one of the three study reservoirs.

the period of spring and summer stratification in Lake Fayetteville. The N<sub>2</sub> accumulation in the hypolimnion was equivalent to a denitrification rate of 62 mg N m<sup>-2</sup> day<sup>-1</sup>. When combined with sediment denitrification rates, the whole-ecosystem denitrification flux in Lake Fayetteville was 17 ± 9 (S.D.) g N m<sup>-2</sup> year<sup>-1</sup>.

Whole-lake N mass balance – An example of whole-lake N mass balance for Lake Fayetteville is shown in Figure 3. We are currently developing mass balance estimates for the entire study period. Nitrogen storage and export was highly dependent upon hydrology. During baseflow conditions the reservoirs were tremendous N sinks. However, the reservoirs acted as net N sources from brief periods during storm events. When considered together, preliminary estimates indicate that the reservoirs store between 10 – 50 g N m<sup>-2</sup> year<sup>-1</sup>.



**Conclusions:** Reservoirs can be substantial N sinks and are hotspots for denitrification. As much as 50 g N m<sup>-2</sup> year<sup>-1</sup> may be trapped by reservoirs, and between 34 – 100% of this stored N is eventually denitrified. These estimates are preliminary and require substantial refinement. However, these preliminary data suggest that reservoirs are very important N sinks and transformers in the landscape that protect downstream water quality by permanently removing reactive N from surface waters.

#### References:

David, M.B., L.G. Wall, T.V. Royer, and J.L. Tank. 2006. Denitrification and the nitrogen budget of a reservoir in an agricultural landscape, *Ecological Applications*, 16: 2177 – 2190

#### Thesis or Dissertation:

Grantz, E. 2011. Denitrification efficiency in water impoundment reservoirs, Department of Crop, Soil, and Environmental Sciences, University of Arkansas.

## **Information Transfer Program Introduction**

Dissemination of information is one of the main objectives of the Arkansas Water Resources Center. To achieve this objective, AWRC sponsors an annual water conference held in Fayetteville, Arkansas, which draws approximately 100 researchers, students, agency personnel, and interested citizens to learn about urgent research and hot topics in water resources throughout the State. AWRC also co-sponsors workshops and other water related conferences in the state and region.

The AWRC maintains a technical library containing over 900 titles, many of which are available online. This valuable resource is utilized by a variety of user groups including researchers students, regulators, planners, lawyers and citizens. Many of the AWRC library holdings have been converted to electronic PDF format which can be accessed via the AWRC website at [www.uark.edu/depts/awrc/publications.htm](http://www.uark.edu/depts/awrc/publications.htm). AWRC is continuing to add archived documents from the library to this electronic data set, and all new titles are added when received.

# Arkansas Water Resources Center Information Transfer Program

## Basic Information

<b>Title:</b>	Arkansas Water Resources Center Information Transfer Program
<b>Project Number:</b>	2010AR245B
<b>Start Date:</b>	3/1/2010
<b>End Date:</b>	2/28/2011
<b>Funding Source:</b>	104B
<b>Congressional District:</b>	3
<b>Research Category:</b>	Not Applicable
<b>Focus Category:</b>	Surface Water, Groundwater, Water Use
<b>Descriptors:</b>	None
<b>Principal Investigators:</b>	Brian E. Haggard

## Publications

1. Massey, L.B., B.E. Haggard, J.M. Galloway, K.A. Loftin, M.T. Meyer, and W.R. Green. 2010. Antibiotic fate and transport in three effluent-dominated Ozark streams. *Ecological Engineering*36(7): 930-938.
2. Haggard, B.E., A.N. Sharply, L.B. Massey and K.M. Teague. 2010. Final Report to the Illinois River Watershed Partnership: Recommended Watershed Based Strategy for the Upper Illinois River Watershed, Northwest Arkansas. Arkansas Water Resources Center, Fayetteville, Arkansas. Miscellaneous Publication 355: 126 pp. In Press.
3. Haggard, B.E., A. Sharpley, and L. Massey eds. 2010. Handbook of Best Management Practices for the Upper Illinois River Watershed and Other Regional Watersheds. Arkansas Water Resources Center, Fayetteville, Arkansas. Miscellaneous Publication 357:120 pp.
4. Pennington, J.H., K.A. Teague, and L.B. Massey. 2010. Management Actions for Developments in Handbook of Best Management Practices for the Upper Illinois River Watershed and Other Regional Watersheds, Haggard, Sharpley and Massey, Eds. Arkansas Water Resources Center, Fayetteville, Arkansas. Miscellaneous Publication 357: 1-18.
5. Pennington, J.H., K.A. Teague, and L.B. Massey. 2010. Management Actions for Municipalities in Handbook of Best Management Practices for the Upper Illinois River Watershed and Other Regional Watersheds, Haggard, Sharpley and Massey, Eds. Arkansas Water Resources Center, Fayetteville, Arkansas. Miscellaneous Publication 357: 19-31.
6. Pennington, J.H., K.A. Teague, and L.B. Massey. 2010. Agricultural Conservation Practices and Programs in Handbook of Best Management Practices for the Upper Illinois River Watershed and Other Regional Watersheds, Haggard, Sharpley and Massey, Eds. Arkansas Water Resources Center, Fayetteville, Arkansas. Miscellaneous Publication 357: 57-82.
7. Haggard, B.E., A.N. Sharpley, and L.B. Massey. 2010. Water Quality and Watershed Conditions in the Upper Illinois River Watershed. Arkansas Water Resources Center, Fayetteville, Arkansas. Miscellaneous Publication 359: 23 pp.
8. Massey, L.B. and B.E. Haggard. 2010. Water Quality Monitoring and Constituent Load Estimation in the Kings River near Berryville, Arkansas, 2009. Arkansas Water Resources Center, Fayetteville, Arkansas. Miscellaneous Publication 361: 16 pp.
9. Massey, L.B., B.E. Haggard, R.S. Avery, and R.A. Morgan. 2010. Water Quality Monitoring and Constituent Load Estimation in the Upper White River Basin, 2009. Arkansas Water Resources

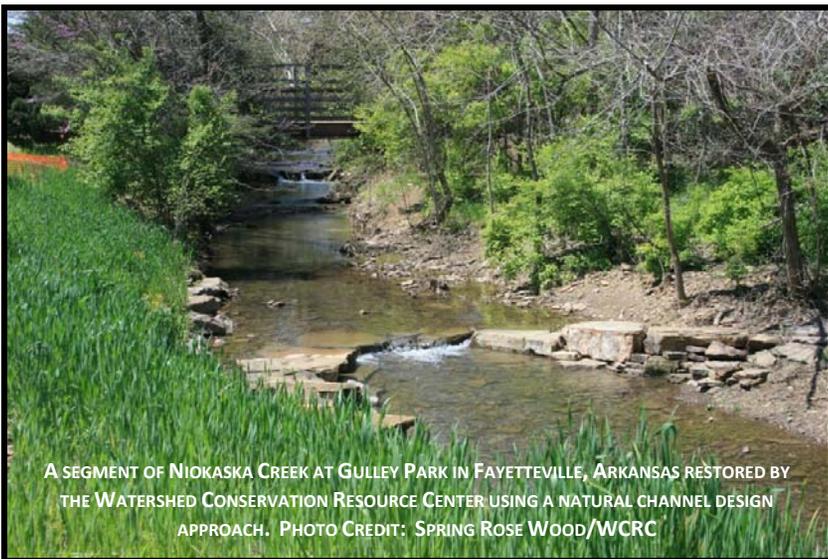
## Arkansas Water Resources Center Information Transfer Program

- Center, Fayetteville, Arkansas. Miscellaneous Publication 362: 40 pp.
10. Massey, L.B. and B.E. Haggard. 2010. Water Quality Monitoring and Constituent Load Estimation in the Upper Illinois River Watershed, 2009. Arkansas Water Resources Center, Fayetteville, Arkansas. Miscellaneous Publication 363: 52 pp.
  11. Haggard, B.E., A.N. Sharpley, L.B. Massey and N. Hardiman. Prioritizing HUC-12 Sub-watersheds using Water Quality Monitoring Data. National Water Quality Monitoring Council, April 25-29, 2010. Denver, Colorado.
  12. Massey, L.B. and B.E. Haggard., 2010. A Volunteer Water Quality Monitoring Program in the Upper Illinois River Watershed, Northwest Arkansas. A Community on Ecosystem Services 2010 Conference, Phoenix, Arizona, December 6-9, 2010.
  13. Haggard, B.E., A.N. Sharpley, L.B. Massey and N. Hardiman. 2010. HUC 12 Watershed Prioritization along a Water Quality and Land Use Gradient: Upper Illinois River Watershed Management Plan. USDA-NIFA 2010 Land Grant and Sea Grant National Water Conference. February 21-25, 2010. Hilton Head, South Carolina.
  14. Hamdan, T.A., T. Scott, D. Wolf, and B.E. Haggard. 2010. Sediment phosphorus flux in Beaver Lake in Northwest Arkansas. *Discovery – The Student Journal of the Dale Bumpers College of Agriculture, Food and Life Sciences*. 11: 3–12.
  15. Washispack, A.N., J.A. McGinnis, and B.E. Haggard. 2010. Assessment of total organic carbon concentrations in two streams of Northwest Arkansas: Town Branch and Brush Creek. *Discovery – The Student Journal of the Dale Bumpers College of Agriculture, Food and Life Sciences*. 11: 51–58.
  16. Migliaccio, K.W., J. Castro, and B.E. Haggard. 2010. Chapter 13 Water Quality Statistical Analysis, Editors: Y. Li and K.W. Migliaccio, *Water Quality Concepts, Sampling, and Analyses*, CRC Press Taylor and Francis Group, LLC. pp. 241-274.
  17. B.E. Haggard and J.T. Scott. 2010. Chapter 3 Water Quality Standards: Designated Uses and Numeric Criteria Development, Editors: Y. Li and K.W. Migliaccio, *Water Quality Concepts, Sampling, and Analyses*, CRC Press Taylor and Francis Group, LLC. pp. 21-40.
  18. Haggard, B.E. 2010. Phosphorus concentrations, loads and sources at the Illinois River, Arkansas, 1997–2008. *Journal of Environmental Quality* 39: 2113-2120.
  19. He, Z., Zhang, H., Toor, G.S., Dou, Z., Honeycutt, C.W., Haggard, B.E., and Reiter, M.S. 2010. Phosphorus distribution in sequentially-extracted fractions of biosolids, poultry litter and granulated products. *Soil Science* 175(4):154-161.
  20. Longing, S.D., and Haggard, B.E. 2010. Distributions of median nutrient and chlorophyll concentrations across the Red River Basin, USA, 1996–2006. *Journal of Environmental Quality* 39: 1966-1974.
  21. Longing, S.D., and Haggard, B.E. 2010. Biological assessment to support ecological recovery of a degraded headwater system. *Environmental Management* 46: 459-470.
  22. Haggard, B., and Scott, T. 2010. Phosphorus release from bottom sediments at Lake Wister, Oklahoma, Summer 2010. Final Report, Poteau Valley Improvement Authority, Poteau, Oklahoma.
  23. Busch, D. and B. Haggard. 2010. Alternative Surface-Water Sampling Methods. UW Platteville, Pioneer Farms: Producer Driven Agricultural Research, 2 pp.
  24. Sharpley, A., P. Moore Jr., K. VanDevender, M. Daniels, W. Delp, B. Haggard, T. Daniels, and A. Baber. 2010. Arkansas Phosphorus Index. UA Division of Agriculture Cooperative Extension Service FSA9531, 8 pp.
  25. Sharpley, A., M. Daniels, K. VanDevender, P. Moore Jr., B. Haggard. N Slaton, and C. West. 2010. Using the 2010 Arkansas Phosphorus Index. UA Division of Agriculture Cooperative Extension Service MP487, 17 pp.



## 2010 ANNUAL RESEARCH AND WATERSHED CONFERENCE

APRIL 13-14, 2010 • FAYETTEVILLE, ARKANSAS  
UNIVERSITY OF ARKANSAS



A SEGMENT OF NIOKASKA CREEK AT GULLEY PARK IN FAYETTEVILLE, ARKANSAS RESTORED BY  
THE WATERSHED CONSERVATION RESOURCE CENTER USING A NATURAL CHANNEL DESIGN  
APPROACH. PHOTO CREDIT: SPRING ROSE WOOD/WCRC

**UofA** UNIVERSITY OF ARKANSAS  
DIVISION OF AGRICULTURE

# Arkansas Water Resources Center

203 Engineering Hall  
University of Arkansas  
Fayetteville, Arkansas 72701

Phone: 479.575.4403  
Email: awrc@uark.edu

[www.uark.edu/depts/awrc](http://www.uark.edu/depts/awrc)

## Brian E. Haggard

Associate Professor and Director  
479.575.2879  
haggard@uark.edu

## Leslie B. Massey

Program Associate and Project Manager  
479.575.2840  
lbartsc@uark.edu

## THE WATER RESOURCES RESEARCH ACT

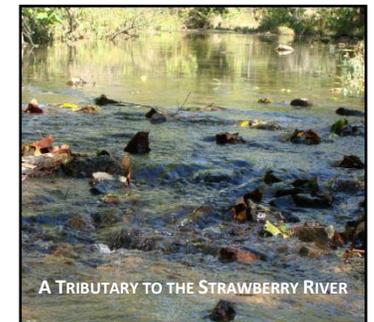


In 1964, Congress passed the Water Resources Research Act, establishing 54 water resources research institutions like Arkansas Water Resources Center at land grant universities throughout the United States. The State Water

Resources Research Center Institutes were charged with arranging for competent research that addresses water problems and enhances our understanding, aiding the entry of new research scientists into water resources fields, helping to train future water scientists and engineers, and transferring results of sponsored research to water managers and the public.

Each fall, the AWRC accepts applications for projects funded through the 104b program. This year, the Arkansas Water Resources Center funded three projects including:

- Determination of the magnitude of mercury methylation in the water column of a high organic carbon river, lower Ouachita River, Union and Ashley Counties, Arkansas, Phil Hays, University of Arkansas and US Geological Survey, \$19,761.
- Assessment of water quality and stream bank stability following BMP implementation on the upper Strawberry River watershed, Jennifer Bouldin, Arkansas State University, \$22,000.
- Denitrification, internal nitrogen cycling and nitrogen retention in river impoundment reservoirs, Thad Scott, UA Division of Agriculture, \$20,579.



# AWRC ANNUAL CONFERENCE PROGRAM AT A GLANCE

**TUESDAY APRIL 13, 2010**

- 7:45-8:15 Coffee, Juice, Pastries  
8:15- 9:30 Session 1: Groundwater Issues near Eureka Springs, Arkansas  
9:30-9:45 Refreshment Break  
9:45-11:35 Session 2: Emerging Issues Facing Groundwater in Arkansas  
11:35-12:30 Lunch Break  
12:30-1:30 Poster Session and Student Poster Competition  
1:30-3:00 Session 3: Reservoirs—Understanding and Managing Ecosystem Services  
3:00-3:15 Refreshment Break  
3:15-5:00 Session 4: Water Supply Reservoirs—Research Needs and Consortium Possibilities  
8:00-9:30 Social at Damgoode Pies

**WEDNESDAY APRIL 14, 2010**

- 7:45- 8:15 Coffee, Juice, Pastries  
8:15- 10:10 Session 5: Improving Aquatic Habitat, Water Quality, and Riparian Areas through Stream Restoration and Bank Stabilization Projects  
10:10-10:30 Coffee Break  
10:30-11:30 Session 6: Improving Aquatic Habitat, Water Quality, and Riparian Areas through Stream Restoration and Bank Stabilization Projects—Continued  
11:55-1:00 Lunch (On Your Own)  
1:00-5:00 Field Tour of West Fork White River Stream Restoration at Brentwood (Meet in front of the Cosmopolitan Hotel)

# ORAL SESSION SCHEDULE

**TUESDAY APRIL 13, 2010**

**SESSION 1: GROUNDWATER ISSUES NEAR EUREKA SPRINGS, ARKANSAS**

**MODERATOR: SUSAN BOLYARD, U.S. GEOLOGICAL SURVEY**

- 8:15** Welcome and Introduction, Brian Haggard, Director, AWRC  
**8:20** The Crumbling Karst of Eureka Springs, Arkansas, Jim Helwig, Consulting Geologist, Eureka Springs (jahelwig@hotmail.com)  
**8:50** Emerging Contaminants Analysis Results for a Single Sampling Event at Eureka Springs, Arkansas—Implications for the Status of Groundwater Quality, Phillip Hays, U.S. Geological Survey (pdhays@usgs.gov)  
**9:10** Results of Initial Reconnaissance of Major Ions, Trace Metals and Nutrients in Groundwater in Eureka Springs, Roger Miller, Arkansas Department of Environmental Quality (millerr@adeq.state.ar.us)  
**9:30** Follow-up Discussion, Questions and Refreshment Break

**SESSION 2: EMERGING ISSUES FACING GROUNDWATER IN ARKANSAS**

**MODERATOR: PHIL HAYS, U.S. GEOLOGICAL SURVEY**

- 9:45** Industry Perspective on Shale Gas Drilling and Production on Water Resources, Doug Melton, Southwest Energy Company, (Doug\_Melton@swn.com) *Key Note Speaker*  
**10:10** Regulatory Perspective on Shale Gas Drilling and Production on Water Resources, Chris Davidson, U.S. Fish and Wildlife Service (chris\_davidson@fws.gov) *Key Note Speaker*  
**10:35** Hot Springs Management Strategy for Recharge Protection, Steve Rudd, National Parks Service (stephen\_rudd@nps.gov)  
**11:00** Surface Water and Groundwater Interaction: Using Groundwater Geochemistry to Assess River-Water Infiltration and Natural geochemical Evolution in the Arkansas river Alluvial Aquifer, Tim Kresse, U.S. Geological Survey (tkresse@usgs.gov)

**LUNCH AND POSTER SESSION**

- 11:35- 12:30** Lunch Break – Penguin Ed’s Barbeque (Foyer and Room 405)  
**12:30- 1:30** Poster Session and Student Poster Competition (Presenters should stand at Posters, Room 405)



## LIST OF POSTERS:

1. Persistence of Broad-Host-Range Plasmids in Municipal Wastewater Treatment Plants Utilizing Chlorination and UV-Irradiation Disinfection Regimes. K.L. Asfahl\*, T. Akiyama and M.C. Savin, Department of Crop, Soil and Environmental Sciences, University of Arkansas, kasfahl@uark.edu.  
*Student Poster Presenter.*
2. Biologically-Mediated Phosphorus Flux in Illinois River Tributaries. B. Drake\* and J.T. Scott, Department of Crop, Soil and Environmental Sciences, University of Arkansas, mdrake@uark.edu.  
*Student Poster Presenter.*
3. Assessment of OHV Trails in the Lee Creek Watershed. P.R. Easley\* and Don Clover, Fort Smith Utility, RandyE@fortsmithar.gov.
4. Tangential Flow, Ultrafiltration and Molecular Detection of Human and Animal Enteric Viruses in Environmental Water Samples. K. Gibson, Johns Hopkins Bloomberg School of Public Health, kgibson@jhsph.edu.
5. Nitrogen Retention and Denitrification Efficiency in Reservoirs. E.M. Grantz\* and J.T. Scott, Department of Crop, Soil and Environmental Sciences, University of Arkansas, egrantz@uark.edu.  
*Student Poster Presenter.*
6. Designing a Demonstration of Nutrient Removal from a Wastewater Treatment Process Utilizing an Algal Growth Bed. J. Hickie and E. Boles. Department of Biological and Agricultural Engineering, University of Arkansas, jphickle@uark.edu, e.boles@uark.edu.  
*Student Poster Presenter.*
7. Modeling Watershed Scale Sediment Loading in the West Fork of the White River using the Soil and Water Assessment Tool. C. N. Jones\*, M.D. Leh, and S.G. Bajwa, Department of Biological and Agricultural Engineering, University of Arkansas, cnj001@uark.edu.  
*Student Poster Presenter.*
8. Application of the Annualized Agricultural Non-Point Source (AnnAGNPS) Model in the West Fork White River Watershed. M.D. Leh\*, S.G. Bajwa and I. Chaubey, Department of Biological and Agricultural Engineering, University of Arkansas, mleh@uark.edu.
9. Cooperative Stakeholder Engagement for Fostering Participation and Gaining Effective Results. J.H. Pennington\*, J. Popp, G. Rodriguez, I. Chaubey, and E. Gbur, Washington County Cooperative Extension, jhpennington@uaex.edu
10. Outreach and Education at Its Finest: Working with all Stakeholder Groups to Address Water Quality Issues in an Impaired Watershed. J.H. Pennington\* and K. Teague, Washington County Cooperative Extension, jhpennington@uaex.edu
11. Sediment-Water Column Interaction for Streams in the Upper Illinois River Watershed. C.W. Rogers\*, A.N. Sharpley, and B.E. Haggard, Department of Crop, Soil and Environmental Sciences, University of Arkansas, cwrogers@uark.edu.  
*Student Poster Presenter.*
12. Effects of Natural Gas Drilling on Stream Quality and Periphyton in the Fayetteville Shale. A. Jackson, M. Evans-White\*, and S. Entekin, Department of Biological Sciences, University of Arkansas, mevanswh@uark.edu.
13. Cooling Broiler Chickens with Less Water Using Controlled Surface Wetting Method. Y. Liang\*, G.T. Tabler, S. Watkins, and I. Berry, Department of Biological and Agricultural Engineering, yliang@uark.edu
14. Connecting Arkansans to their Watershed. S. DeVries\* and K. Finefield, Arkansas Watershed Advisory Group, DEVRIES@adeq.state.ar.us.





## DAMGOODE PIES SOCIAL

Tuesday, April 13 from 8:00-9:30 PM  
Sponsored by Hach Environmental

Damgoode Pies is located at 37 East Center Street  
(Just East of the Square)  
~ 2 minute walk from the Cosmopolitan Hotel



HACH Hydromet manufactures Hydrolab water quality instruments and OTT water level/discharge instruments. Contact Dave Procyk, Factory Direct Manager, at 512-288-5831 or check them out at [www.hachhydromet.com](http://www.hachhydromet.com).



## WEDNESDAY APRIL 14, 2010

### SESSION 5: IMPROVING AQUATIC HABITAT, WATER QUALITY, AND RIPARIAN AREAS THROUGH STREAM RESTORATION AND BANK STABILIZATION PROJECTS MODERATOR: MAT VAN EPPS, WATERSHED CONSERVATION RESOURCE CENTER

- 8:15** Stream Channel Process and Ecological Services, Eric Cummings, UA Division of Agriculture ([ecummin@uark.edu](mailto:ecummin@uark.edu))
- 8:40** Prioritizing Stream Reaches for Restoration on the West Fork White River, Sandi Formica, Watershed Conservation Resource Center ([formica@watershedconservation.org](mailto:formica@watershedconservation.org))
- 9:10** Designing and Constructing Stream Restoration to Improve Habitat for Fish and Mussel Species of Concern in the Saline River, Joy DeClerk, The Nature Conservancy ([jdeclerk@tnc.org](mailto:jdeclerk@tnc.org))
- 9:40** Improving Arkansas Fisheries and Providing Hands-On Environmental Education through Streambank Restoration Projects, Dave Evans and Tim Burnley, Arkansas Game & Fish Commission ([daevans@agfc.state.ar.us](mailto:daevans@agfc.state.ar.us); [tburnley@agfc.state.ar.us](mailto:tburnley@agfc.state.ar.us))
- 10:10** Refreshment Break

### SESSION 6: IMPROVING AQUATIC HABITAT, WATER QUALITY, AND RIPARIAN AREAS THROUGH STREAM RESTORATION AND BANK STABILIZATION PROJECTS—CONTINUED MODERATOR: MARTY MATLOCK, CENTER FOR AGRICULTURAL AND RURAL SUSTAINABILITY

- 10:30** Addressing Headwater Streams as a Sediment Source in the Delta, Matt Lindsey, The Nature Conservancy ([mlindsey@tnc.org](mailto:mlindsey@tnc.org))
- 11:00** Design and Construction of a Streambank Stabilization Project to Improve Habitat for the Yellowcheek Darter and Speckled Pocketbook Mussel in the Little Red River Watershed, Josh Duzan and Ethan Inlander, The Nature Conservancy ([jduzan@tnc.org](mailto:jduzan@tnc.org); [einlander@tnc.org](mailto:einlander@tnc.org))
- 11:30** Design and Implementation of Urban and Rural Stream Restoration to Reduce Sediment and Phosphorus Loadings in the Illinois River and Beaver Lake Watersheds, Matt Van Eps, Watershed Conservation Resource Center ([vaneps@watershedconservation.org](mailto:vaneps@watershedconservation.org))
- 11:55** Lunch Break (On Your Own)





## PRE-REGISTERED CONFERENCE PARTICIPANTS

Bodie Drake  
UA Division of Agriculture  
bmdrake@uark.edu

Josh Duzan  
The Nature Conservancy  
jduzan@tnc.org

Randy Easley  
Fort Smith Utility  
RandyE@fortsmithar.gov

Dave Evans  
Arkansas Game & Fish Commission  
daevans@agfc.state.ar.us

Michelle Evans-White  
University of Arkansas  
mevanwh@uark.edu

Kate Finefield  
ADEQ/AWAG  
finefield@adeq.ar.us

John Fohner  
University of Arkansas  
jfohner@uark.edu

Sandi Formica  
Watershed Conservation and Resource Center  
formica@watershedconservation.org

Alan Fortenberry  
Beaver Water District  
afortenberry@bwdh2o.org

Dave Freiwald  
US Geological Survey  
freiwald@usgs.gov

Colene Gaston  
Beaver Water District  
cgaston@bwdh2o.org

Kristen Gibson  
Johns Hopkins Bloomberg School  
kgibson@jhspsh.edu

Erin Grantz  
University of Arkansas  
egrantz@uark.edu

Reed Green  
US Geological Survey  
wrgreen@usgs.gov

Brian Haggard  
UA Division of Agriculture  
haggard@uark.edu

Robert Hart  
Arkansas Department of Health  
Robert.hart@arkansas.gov

Phil Hays  
UA/US Geological Survey  
pdhays@uak.edu

Jim Helwig  
Consulting Geologist  
jahelwig@hotmail.com

Jeff Hickle  
University of Arkansas  
jphickle@uark.edu

Ethan Inlander  
The Nature Conservancy  
einlander@tnc.org

Janis Jones  
US Army Corps of Engineers  
jan.r.jones@usace.army.mil

Nathan Jones  
University of Arkansas  
cnj001@uark.edu

## PRE-REGISTERED CONFERENCE PARTICIPANTS

Evelyn Kort  
AR Dept. of Environmental Quality  
kort@adeq.state.ar.us

Tim Kresse  
US Geological Survey  
tkresse@usgs.gov

Hal Liechty  
UA Monticello  
liechty@uamont.edu

Mansoor Leh  
UA Division of Agriculture  
mleh@uark.edu

Matt Lindsey  
The Nature Conservancy  
mlindsey@tnc.org

Larry Lloyd  
Beaver Water District  
llloyd@bwdh2o.org

Martin Maner  
Central Arkansas Water  
Martin.maner@carkw.com

Leslie Massey  
Arkansas Water Resources Center  
lbartsc@uark.edu

Doug Melton  
Southwestern Energy  
doug\_melton@swn.com

Roger Miller  
AR Dept. of Environmental Quality  
millerr@adeq.state.ar.us

Richard Monk

Bob Morgan  
Beaver Water District  
rmorgan@bwdh2o.org

Craig Noble  
Russellville Water Utilities  
cnoble@citycorporation.com

Steve Patterson  
Bio x Design  
spatterson5000@earthlink.net

John Payne  
UA Division of Agriculture  
jpayne@uaex.edu

John Pennington  
UA Cooperative Extension Service  
jhpennington@uaex.edu

Greg Phillips  
GBMc & Associates  
gphillips@gbmcassoc.com

David Procyk  
HACH Hydromet  
dprocyk@hach.com

Patrick Pruitt  
Rogers Water Utilities  
Xpatrickpruitt@rwu.org

Andrea Radwell  
University of Arkansas  
aradwell@uark.edu

Steve Ricke  
University of Arkansas  
sricke@uark.edu

Tom Riley  
UA Division of Agriculture  
triley@uaex.edu

## PRE-REGISTERED CONFERENCE PARTICIPANTS

Chris Rogers  
UA Division of Agriculture  
cwrogers@uark.edu

Steve Rudd  
National Parks Service  
stephen\_rudd@nps.gov

Randy Rushin  
Water Monitoring Solutions, Inc.  
randy@water-monitor.com

Thad Scott  
UA Division of Agriculture  
jts004@uark.edu

Stan Starling  
Arkansas Department of Health  
stanley.starling@arkansas.gov

Rusty Tate  
Beaver Water District  
rtate@bwdh2o.org

Katie Teague  
UA Cooperative Extension Service  
kteague@uaex.edu

Kent Thornton  
FTN Associates, Ltd.  
kwt@ftn-assoc.com

Matt Van Eps  
Watershed Conservation Resource Center  
vaneps@watershedconservation.org

Renee Vardy  
University of Arkansas  
rvardy@uark.edu

Len Weeks  
USDA Forest Service  
lweeks@fs.fed.us

Amy Wilson  
Beaver Water District  
awilson@bwdh2o.org

## CONFERENCE SPONSORS AND EXHIBITORS

Mark Cochran  
Associate Vice President  
for Agriculture- Research  
Fayetteville, Arkansas

Dave Procyk  
Regional Manager  
Austin, Texas

Amy Wilson  
Director of Public Affairs  
Lowell, Arkansas

Randy Rushin  
Water Monitoring Solutions, Inc.  
Sulphur Springs, Texas

David Freiwald  
Arkansas Water Science Center  
Little Rock, Arkansas

Sarah DeVries  
AWAG Coordinator  
North Little Rock, Arkansas



# USGS Summer Intern Program

None.

<b>Student Support</b>					
<b>Category</b>	<b>Section 104 Base Grant</b>	<b>Section 104 NCGP Award</b>	<b>NIWR-USGS Internship</b>	<b>Supplemental Awards</b>	<b>Total</b>
<b>Undergraduate</b>	4	0	0	3	7
<b>Masters</b>	2	0	0	2	4
<b>Ph.D.</b>	3	0	0	0	3
<b>Post-Doc.</b>	0	0	0	0	0
<b>Total</b>	9	0	0	5	14

## **Notable Awards and Achievements**

Brian Haggard, Director of the Arkansas Water Resources center received the College of Engineering Outstanding Researcher award for the department of Biological and Agricultural Engineering at the University of Arkansas, 2010-2011.