

# **Report for 2005NC49B: A Comparison of Drought Tolerance in Common Herbaceous Wetland Macrophytes as Indicated by Plant Growth, Water Status, and Oxidative Stress**

## Publications

- Articles in Refereed Scientific Journals:
  - Touchette, B.W., In Press, Salt tolerance in a *Juncus roemerianus* brackish marsh: Spatial variations in plant water relations. *Journal of Experimental Marine Biology and Ecology*.

Report Follows

## Title

A Comparison of Drought Tolerance in Common Herbaceous Wetland Macrophytes as Indicated by Plant Growth, Water Status, and Oxidative Stress (70216)

## Problem and Research Objectives

### **Part 1: Drought Susceptibility in Emergent Wetland Angiosperms: a Comparison of Water Deficit Growth in Five Herbaceous Perennials**

Wetland ecosystems by nature are integrally tied to hydrology (Collins and Battaglia 2001; De Steven and Toner 2004). Consequently, mechanisms that alter prevailing hydrologic conditions, including mechanical and climatological, may sharply influence wetland function and value. Whereas water fluctuations involving episodic inundations and desiccation can facilitate increased plant diversity (Bush et al. 1998; Olson 2004; Mulhouse et al. 2005), extended periods of submersion or drought may foster decreased plant diversity, including the possible promotion of monotypic stands of invasive aquatic species (Pezeshki et al. 1998; Galatowitsch et al. 2000; Bonilla-Warford and Zedler 2002; Kercher and Zedler 2004). Williams and Hudak (2005) suggested that drought and low water availability are among the greatest threats to constructed wetlands in north-central Texas, and that herbaceous species (e.g., *Sagittaria latifolia*, and *Eleocharis quadrangulata*) are particularly vulnerable to water deficits. Similarly, Holland et al. (1995) reported the loss of several urban wetlands in Oregon due to drought-associated water deficits, and that drier-end wetlands (e.g., seasonally flooded) were often more susceptible to human disturbances. Moreover, droughts beyond recoverable time periods for established macrophytes, could eliminate valuable habitat necessary for both migratory waterfowl and indigenous wildlife (Sorenson et al. 1998; Williams and Hudak 2005).

Despite selective environmental pressures that would seemingly promote tolerance to both flooding and exposure, many wetland plants are unable to withstand even short periods of water scarcity (Stuedler and Touchette 2003; Mulhouse et al. 2005; Williams and Hudak 2005). This is a fundamental concern, as wetlands are considered to be among the most threatened habitats globally (Gopal and Junk 2000). In a paper published in 2002, Jacobs et al. noted our lack of knowledge and the need for further studies on how extreme water table fluctuations influence wetland vegetation. While, there is growing interest in defining drought-associated changes in natural vegetation, driven primarily by climate change models predicting increases in drought severity and duration in the mid-latitudes, much of this work has been focused on grasslands and forests (e.g., Mangan et al. 2004; Breshears et al. 2005), and comparatively fewer studies have exclusively considered wetland systems (Sorenson et al. 1998; Dawson et al. 2003). Therefore, the purpose of this study was to evaluate growth responses of five herbaceous wetland species (monocots *Carex alata*, *Juncus effusus*, and *Peltandra virginica*, and dicots *Saururus cernuus*, and *Justicia americana*) to simulated drought conditions (up to 6 wks with 1-in-25 yr precipitation low and water withdrawal). Emergent herbaceous plants were selected in favor of woody vegetation because of their ease of manipulation under

controlled greenhouse conditions and due to their relative vulnerability to low water supply (Williams and Hudak 2005). For this study, it was anticipated that the herbaceous wetland plants would demonstrate varying degrees of drought tolerance, ranging from rapid vegetative die-off to minimal adverse effects. Thereby, the results of this study could be useful in providing a framework to gauge relative tolerance to drought among different wetland plant species. As a secondary application, this study could also provide valuable insight into proper plant selection for wetland restoration/ creation in areas with sporadic water availability (including stormwater retention wetlands).

Freshwater wetlands often exist as transitional areas between terrestrial uplands and deep open water. Thus they are fundamentally sensitive to changes in hydrology. Some of the more dramatic changes in wetland water supply occur during extensive droughts, where both precipitation and soil water table markedly decline. While it is generally understood that herbaceous wetland macrophytes are more sensitive to decreased water availability than wetland trees, the degree of susceptibility among wetland herbs remains relatively unexplored. Therefore, the purpose of this study was to evaluate plant growth responses of five herbaceous wetland species (monocots *Carex alata*, *Juncus effusus*, and *Peltandra virginica*, and dicots *Saururus cernuus*, and *Justicia americana*) to simulated drought conditions (up to 6 wks in a 1-in-25 yr precipitation low with receding soil water tables).

## **Part 2: Drought Tolerance Versus Drought Avoidance: A Comparison of Plant-Water Relations in Herbaceous Wetland Plants Subjected to Water Withdrawal and Repletion**

Water deficits can result in significant declines in overall plant productivity and with increasing water scarcity can promote high rates of plant mortality. While a number of studies have addressed physiological acclimations to low water availability on agriculturally important species, comparatively fewer studies have considered wetland vegetation. This discrepancy is not surprising when considering aquatic plants, by nature, are constrained to environments characteristically dominated by excessive water. Nevertheless, wetlands represent an intermediate between terrestrial and open-water systems, and are thus fundamentally sensitive to changes in hydrology. Seemingly slight changes in water level may result in substantial adjustments in both species richness and productivity (Mitch and Gosselink 1986; Amlin and Rood 2002). Furthermore, when considering wetland hydroperiods, some systems (e.g., intermittently exposed, and seasonally flooded wetlands) can sustain extended periods of flooding and exposure (Cowardin et al. 1979; Wilcox 2002). In extreme cases, seasonally flooded wetlands will undergo enormous fluctuations in water availability as soil water tables oscillate from 50 cm aboveground to 100 cm belowground over a period of weeks (Mitsch et al. 1979). Furthermore, the degree of water deprivation can worsen during episodes of unseasonably low precipitation or drought; influencing both the degree of water deficit and extending the period of which these aquatic plants must tolerate water stressed conditions.

Drought, as defined as the absence of appreciable precipitation over an extended period of time allowing for the depletion of soil moisture with a concomitant injury to plants

(Kramer 1983), can influence natural plant distributions and productivity. Adjustments by plants to drought may involve avoidance and/or tolerance. Drought avoidance includes responses such as increased stomatal and cuticular resistance, changes in leaf area and anatomy, and changes in leaf orientation (Morgan 1984; Jones and Corlett 1992; Zlatev 2005). Whereas, drought tolerance involves maintaining adequate cell turgor, while preventing disruptions in cellular metabolism (Munns 1988; Savé et al. 1993). Tolerance has been attributed to at least two mechanisms - osmotic adjustment (involving inorganic ions, carbohydrates, and organic acids), and changes in cellular/ tissue elasticity (i.e., bulk elastic modulus;  $\epsilon$ ).

Studies suggest that plant metabolic processes are more responsive to turgor and cell volume conditions rather than fluctuations in water potential (Jones and Corlett 1992; Zlatev 2005). While dehydration of cells during water deficits can result in lower osmotic potential by confining existing solutes into smaller volumes, true osmotic adjustment necessitates the accumulation and buildup of these ions or compounds in excess of pre-stress conditions (Bray 1993). Drought-induced changes in tissue elasticity can also modify the relationship between turgor pressure and cell volume contributing further to drought tolerance (Blake et al. 1991; Saito and Terashima 2004).

Understanding the basis of plant water relations in emergent wetland herbs may provide some insight into the capacity of wetland species to maintain metabolic activity during extended periods of drought. Furthermore, any species that is capable of modifying their water relations during periods of low water availability may have a competitive advantage over plants incapable of altering water status. Therefore, the purpose of this study was to evaluate the effects of water stress on plant water relations in five herbaceous wetland species (monocots *Carex alata* Torr., *Juncus effusus* L., *Peltandra virginica* L., and dicots *Saururus cernuus* L., *Justicia americana* L. Vahl.). The ability of these plants to adjust to simulated drought conditions was evaluated using pressure-volume isotherms, which provided insights into leaf osmotic adjustment, tissue elasticity, plant water potential, and turgor pressure. Furthermore, plant productivity and water use efficiencies (WUE) were also evaluated to gain additional insights into wetland plant responses associated with short periods of water deficit and repletion.

## **Methodology**

### **Part 1: Drought Susceptibility in Emergent Wetland Angiosperms: a Comparison of Water Deficit Growth in Five Herbaceous Perennials**

#### *Growth Conditions*

Five herbaceous obligate wetland plants (*Carex alata* Torr., *Juncus effusus* L., *Peltandra virginica* L., *Saururus cernuus* L., and *Justicia americana* L. Vahl.) were grown under controlled greenhouse conditions in 20 L microcosms containing natural lake water (~ 25 cm) and sediments (~20 cm) at plant densities comparable to those observed in natural field populations (60-100 shoots m<sup>-2</sup>). Prior to the initiation of water deficits, the plants were allowed to acclimate within the microcosms for 4 weeks to ensure plant establishment as indicated by active growth and productivity. Drought conditions were

initiated by removing surface water, over a 1-wk period, until the water level reached the upper portion of the substratum. During this period drought-treated plants (n=5 for each treatment) received watering of foliage and sediment comparable to a 1-in-25 year low precipitation rate and periodicity (1.0 cm water at 7-day intervals; based on summer precipitation data from the central Piedmont region of North Carolina over a 75 yr period). Control microcosms (n=5) were watered at mean summer precipitation rates and periodicity (2.9 cm water at 7-day intervals), while maintaining water levels at 25 cm above the substratum. Drought treated microcosms were exposed to 2-, 4-, or 6-wks of simulated drought, followed by a 2- to 6-wk recovery period consisting of pre-drought conditions (i.e., mean summer precipitation rates and periodicity, and 25 cm of standing water). Throughout the study, temperature (daily maximum and minimum) and relative humidity (RH; daily maximum and minimum) were monitored using a temperature and humidity data logger (Hobo H8 logger, Onset Computer Corporation, Bourne, MA).

#### *Growth and Productivity Measurements*

Growth measurements (n = 5) were recorded at weekly intervals and included phytomass (above- and belowground tissue), leaf area (LA), relative growth rate (RGR), and unit leaf rate (ULR; also termed net assimilation rate). Phytomass was determined by carefully removing plants from microcosms, as facilitated by the soft sediments which minimized loss of belowground tissues. The plants were sorted between aboveground- (stems and leaves) and belowground tissues (roots and rhizomes), and dried (70°C) to constant weight. LAs were measured using scanned digital images (7100 USB scanner, Visioneer Inc., Pleasanton, CA) of individual leaves against a white background (as described in Ferris et al. 2001, with modifications described in O'Neal et al. 2002). The images were then imported into an image processing and analysis program, Scion Image (Scion Corporation, Frederick, MD), for leaf area calculations. RGRs were calculated based on the production of dry matter at weekly intervals as defined by the following equation:

$$\text{RGR} = \frac{\ln W_{x+1} - \ln W_x}{t_{x+1} - t_x}$$

where W is the weight of dry matter reported for consecutive collection periods (x and x+1), and t represents the time interval (in wks) between collections. While RGRs are considered a fundamental measure of plant productivity and are important in the comparisons of plant performance between species and/or treatment effects, it is limited in its ability to evaluate causal factors that shape plant productivity (Beadle 1985). Differences in productivity are often associated with the plant's assimilation capacity as reflected in leaf area. Therefore, ULR measurements are often considered in growth analyses because they encompass both dry-mass production and assimilation capacity. ULR is defined as the increase in plant dry-mass over time per unit of assimilatory material, and was calculated in this study as follows:

$$(W_{x+1} - W_x) (\ln LA_{x+1} - \ln LA_x)$$

$$\text{ULR} = \frac{\text{LA}_{x+1} - \text{LA}_x}{(t_{x+1} - t_x)}$$

where LA (as described above) and W were measured at consecutive sample periods (Beadle 1985; Hunt 1990).

#### *Data Analysis*

Along with other measurements, mean percent differences from controls for growth parameters were presented for comparisons. These values were calculated as follows:

$$\% \text{ Difference} = \left( 1 - \frac{\text{treatment}}{\text{control}} \right) * 100$$

and allowed for general evaluations between treatments and their respective controls. These values were not analyzed statistically, and were used to merely represent the magnitude of change when significant differences were observed.

A repeated measure ANOVAs (general linear model [GLM] procedure) were performed using SAS statistical software (SAS Institute Inc., Cary, NC) on RGR, ULR, phytomass, leaf area, and aboveground/belowground tissue ratios. Pre-planned comparisons were performed for each sample collection using LS-means to evaluate treatment responses at each time interval. All comparisons were considered statistically significant when p-values were less than 0.05.

## **Part 2: Drought Tolerance Versus Drought Avoidance: A Comparison of Plant-Water Relations in Herbaceous Wetland Plants Subjected to Water Withdrawal and Repletion**

### *Growth Conditions*

Five herbaceous obligate wetland plants (*Carex alata* Torr., *Juncus effusus* L., *Peltandra virginica* L., *Saururus cernuus* L., and *Justicia americana* L. Vahl.) were grown under controlled greenhouse conditions in 20 L microcosms containing natural lake water (~ 25 cm) and sediments (~20 cm) at plant densities comparable to those observed in natural field populations (60-100 shoots m<sup>-2</sup>). Prior to the initiation of water deficits, the plants were allowed to acclimate within the microcosms for 4 weeks to ensure plant establishment as indicated by active growth and productivity. Drought conditions were initiated by removing surface water, over a 1-wk period, until the water level reached the upper portion of the substratum. During this period drought-treated plants (n=5 for each treatment) received watering of foliage and sediment comparable to a 1-in-25 year low precipitation rate and periodicity (1.0 cm water at 7-day intervals; based on summer precipitation data from the central Piedmont region of North Carolina over a 75 yr period). Control microcosms (n=5) were watered at mean summer rates and periodicity (2.9 cm water at 7-day intervals), while maintaining water levels at 25 cm above the substratum. Drought treated microcosms were exposed to 2 wks of simulated drought, whereas drought-recovered microcosms consisted of 2 wks of simulated drought

followed by a 2 wks return to pre-drought conditions (i.e., mean summer precipitation rates and periodicity, and 25 cm of standing water).

#### *Plant-Water Status*

Leaf relative water content ( $\theta$ ) was evaluated according to Joly (1985) using the following equation:

$$\theta = \frac{(W_f - W_d)}{(W_t - W_d)}$$

where  $W_f$  was the fresh weight recorded during collection,  $W_t$  was the turgid weight, and  $W_d$  was the oven dry weight (70°C, until constant weight). Pre-dawn  $\Psi_{\text{xylem}}$  were determined by using a Scholander pressure chamber (Model 1000, PMS Instrument Co., Albany, Oregon, USA) on young fully extended leaves enclosed within foil bags for 2 hrs.

A Scholander pressure chamber was used to determine  $\Psi_{\text{leaf}}$  on leaves from each species to compare water status of control and drought recovered plants. Water deficits were established by exposing leaves to transpirational water loss on a laboratory bench. Turner et al. (1984) favored this approach, over elevated pressurization, because it minimized the possibility of  $\Psi$  disequilibria between apoplastic and symplastic tissues. Pressure-volume isotherms were constructed by plotting the reciprocal of  $\Psi_{\text{leaf}}$  against  $\theta$ . First order regression analyses were performed on the linear portion of the curve. This line is equivalent to leaf  $\Psi_{\pi}$ , and can be used to determine  $\theta$  at turgor loss point ( $\theta^{\text{tlp}}$ ), the osmotic potential at full saturation ( $\Psi_{\pi}^{\text{sat}}$ ), osmotic potential at turgor loss point ( $\Psi_{\pi}^{\text{tlp}}$ ), and symplastic volume of the total water content ( $\theta_{\text{sym}}$ ). Bulk elastic modulus ( $\epsilon$ ) was obtained from the initial part of the curve, following  $\Psi_{\pi}$  correction, as described in the following equation:

$$\epsilon = \frac{d \Psi_p}{d \theta} \theta_{\text{sym}}$$

where changes in turgor potential ( $\Psi_p$ ) were compared against changes in  $\theta$ , and relative symplastic water content (Koide et al. 1989).

Data collected from pressure-volume isotherms were also used to generate Höfler diagrams for control and drought accumulated plants. The values derived for each point within the diagram represents the mean value among replicates with comparable water content. Because  $\theta$  for these replicates were within ranges, the best polynomial fit represents a population estimate for the mean value within that range of water content. Consequently, data represented by Höfler diagrams may not precisely match the data derived from P-V analysis on each individual plant.

#### *Transpiration and Water Use Efficiency*

A gravimetric technique was used to estimate transpiration on whole plants according to Slavík (1974) and Kramer (1983). This involved careful removal of plants from the

microcosm and enclosing the sediment/ root complex in polyethylene bags to prevent water evaporation from the soil. The soft sediment used during this experiment facilitated the plants removal with negligible damage to root structure. The plants were maintained within a greenhouse under environmental conditions comparable to plants within the microcosms (i.e., 10% neutral density shading of ambient light;  $26 \pm 2^\circ\text{C}$ ;  $45.4 \pm 3.2\%$  RH). Transpiration measurements were integrated over the course of the day with measurements reported at 60 min intervals. Drought treated plants were restricted to sediments collected from the 2-week drought treatment with no additional water added, whereas control and drought-recovered plants were placed in saturated soils with standing water to replicate typical water supply.

Water use efficiency (WUE) was estimated according to Kramer (1983; as modified by Gaiser et al. 2004, and Gao et al. 2004), defined as the amount of water used (via transpiration) per unit dry matter produced.

$$\text{WUE} = \frac{\text{Water used in transpiration}}{\text{Dry matter produced}}$$

Dry matter productivities were estimated as the mean increase in total biomass (both aboveground and belowground tissues) reported during the week of transpiration measurements ( $n = 5$  for each microcosm).

#### *Growth and Productivity*

Growth measurements included both absolute growth (i.e., the amount of dry matter produced  $\text{plant}^{-1} \text{day}^{-1}$ ), and relative growth (i.e., the amount of dry matter produced  $\text{gram}^{-1} [\text{of plant}] \text{day}^{-1}$ ). In this case plants were collected at weekly intervals ( $n=5$ ) and the increase in total dry mass (above- and belowground biomass collectively) were used to estimate plant productivity/ growth. Survival measurements were estimated as the percent change in living plants over the two week drought treatment.

#### *Data Analysis*

Data for controls and drought treated plants were compared using a non-parametric Kruskal-Wallis one-way analysis of variance (ANOVA), followed by a Tukey multiple comparison test for post-hoc evaluations. All comparisons were considered significant at a p-value of less than 0.05.

### **Principal Findings**

#### **Report 1: Drought Susceptibility in Emergent Wetland Angiosperms: a Comparison of Water Deficit Growth in Five Herbaceous Perennials**

Of the five species studied, three (*J. americana*, *S. cernuus*, and *J. effusus*) had no survivors after six weeks of simulated drought. *J. americana*, appeared to be the most sensitive to water deprivation with a 67% decrease in plant phytomass and an 85% decrease in leaf area with only two weeks of drought, and complete mortality after three

weeks. While *P. virginica* also had significant decreases in biomass, leaf area, relative growth rate (RGR) and unit leaf rate (ULR), in as little as two weeks of drought, no noticeable decreases in survival were observed. In contrast, when *J. effusus* experienced between 2- and 4-weeks of water deprivation, there were significant increases in RGR, ULR, phytomass, leaf area, and shoot: root ratios. *S. cernuus* and *C. alata* remained relatively unaffected following four weeks of drought; however by the fifth week, there were significant declines in leaf area for both species. In general, this study provides experimental evidence on how herbaceous macrophytes grow under drought conditions. This basic understanding is fundamental if we are to develop better working models on how wetlands will respond to changing environmental conditions that lead to decreased water supply.

In conclusion, our current understanding on how emergent wetland perennials respond to water deficits is incomplete. This study provides experimental evidence on how plant growth and productivity can be altered during drought events. In general, plants in this study were severely impacted (*J. americana*), moderately impacted (*S. cernuus*, and *P. virginica*), largely unaffected (*C. alata*), or benefited (*J. effusus*) by a combination of decreased precipitation and water table drawdown that simulated a 1-in-25 year drought.

**Table 2.** Summary of growth responses observed in five species of herbaceous wetland plants to 2-, 4-, and 6-wks of drought. Parameters include relative growth rate (RGR), unit leaf rate (ULR), total plant phytomass (Biomass), leaf area, shoot: root ratio (S:R ratio), and percent survival. Numerical values represent the mean maximum percent difference observed from controlled (emergent) conditions, and the values in parentheses represent the week when the greatest difference was observed. The plus or minus signs indicate an increase/ positive response or a decrease/ negative response. Survival is based on the number of individuals remaining as a percent of the initial (for weeks 4 and 10). NC was used to indicate no significant change from the control (repeated measures ANOVA).

Species	RGR	ULR	Biomass	Leaf Area	S:R Ratio	% Survival	
						Week-4	Week-10
<i>Justicia americana</i>							
2-wk	NC	NC	- 67 <sup>(7)</sup>	- 85 <sup>(7)</sup>	NC	75 ± 15	87 ± 7
4-wk <sup>5</sup>	NC	NC	- 46 <sup>(5)</sup>	- 19 <sup>(5)</sup>	+ 83 <sup>(5)</sup>	61 ± 22	0 ± 0
6-wk <sup>5</sup>	NC	NC	- 48 <sup>(5)</sup>	- 35 <sup>(5)</sup>	NC	54 ± 8	0 ± 0
<i>Saururus cernuus</i>							
2-wk	NC	NC	NC	NC	+ 369 <sup>(7)</sup>	84 ± 19	80 ± 18
4-wk	NC	NC	NC	NC	+ 351 <sup>(5)</sup>	83 ± 24	86 ± 15
6-wk <sup>7</sup>	NC	NC	- 75 <sup>(7)</sup>	-32 <sup>(7)</sup>	NC	32 ± 8	0 ± 0
<i>Peltandra virginica</i>							
2-wk	- 231 <sup>(4)</sup>	- 235 <sup>(4)</sup>	- 53 <sup>(10)</sup>	- 64 <sup>(5)</sup>	NC	91 ± 7	103 ± 10
4-wk	- 395 <sup>(6)</sup>	- 394 <sup>(6)</sup>	- 64 <sup>(8)</sup>	+ 44 <sup>(3)</sup> / <sup>-77</sup> (8)	NC	90 ± 7	98 ± 4
6-wk	- 226 <sup>(6)</sup>	- 356 <sup>(7)</sup>	- 84 <sup>(9)</sup>	- 82 <sup>(9)</sup>	NC	93 ± 13	94 ± 9
<i>Juncus effusus</i>							
2-wk	+1062 <sup>(4)</sup>	+ 1579 <sup>(4)</sup>	+ 306 <sup>(10)</sup>	+ 180 <sup>(4)</sup>	NC	86 ± 18	92 ± 9
4-wk	+692 <sup>(4)</sup>	+ 842 <sup>(4)</sup>	+ 198 <sup>(10)</sup>	+ 136 <sup>(10)</sup>	NC	115 ± 30	96 ± 24
6-wk <sup>8</sup>	+655 <sup>(4)</sup>	+ 925 <sup>(4)</sup>	NC	+175 <sup>(4)</sup>	+ 209 <sup>(8)</sup>	49 ± 4	0 ± 0
<i>Carex alata</i>							
2-wk	NC	NC	NC	NC	NC	113 ± 9	108 ± 13
4-wk	NC	NC	NC	NC	NC	126 ± 22	122 ± 25
6-wk	NC	NC	NC	-47 <sup>(10)</sup>	NC	98 ± 3	79 ± 4

## **Part 2: Drought Tolerance Versus Drought Avoidance: A Comparison of Plant-Water Relations in Herbaceous Wetland Plants Subjected to Water Withdrawal and Repletion**

In this study, simulated drought resulted in significant decreases in xylem water potential ( $\Psi_{\text{xylem}}$ ) for all five species, suggesting that these plants were physiologically affected by water deficit. Four of the five species showed outward signs of drought avoidance, including significant reductions in transpiration (*C. alata*, *P. virginica*, *J. americana*, and *S. cernuus*) and modifications of leaf area (*P. virginica* and *J. americana*). Interestingly, while adjustments in transpiration were observed for most plants during the dry period, no significant changes in water use efficiencies (WUE) were detected until after water repletion. That is, only two species (*C. alata* and *P. virginica*) had enhanced WUE as water availability returned to normal. Drought conditions also promoted drought tolerance responses in all five species, as indicated by a change in bulk modulus of elasticity ( $\epsilon$ ; all species) and decreased osmotic potential ( $\Psi_{\pi}$ ; *P. virginica*). Taken as a whole, this study reveals two contrasting drought tolerance strategies in wetland herbs. While four of the species alter  $\epsilon$  to generate declines in  $\Psi$ , *P. virginica* favored decreases in osmotic potentials (as indicated by decreases in  $\Psi_{\pi}$  at full saturation and at turgor loss point).

**Table 1.** Plant and soil parameters collected following two weeks of simulated drought. Data includes relative water content ( $\theta$ ), xylem water potential ( $\Psi_{\text{xylem}}$ ; MPa), aboveground biomass, belowground biomass, and percent survival. Letters represent the statistical relationship between each treatment when differences were detected (the same letter on different values are considered statistically similar). Data is presented as means  $\pm$  1 S.E. (n = 5).

Species (treatment)	$\theta$ (%)	$-\Psi_{\text{xylem}}$	Aboveground (g)	Belowground (g)	Survival (%)
<u>Monocots</u>					
<i>Carex alata</i>					
Control	91.7 $\pm$ 3.7	0.24 $\pm$ 0.07	0.03 $\pm$ 0.003	0.007 $\pm$ 0.001	108.3 $\pm$ 8.3
Drought	84.5 $\pm$ 7.3	0.76 $\pm$ 0.18*	0.04 $\pm$ 0.009	0.004 $\pm$ 0.004	125.5 $\pm$ 21.7
Recovered	91.1 $\pm$ 2.4	0.51 $\pm$ 0.07*	0.03 $\pm$ 0.004	0.004 $\pm$ 0.003	113.3 $\pm$ 9.4
<i>Juncus effusus</i>					
Control	91.2 $\pm$ 3.3	0.66 $\pm$ 0.08	0.14 $\pm$ 0.01	0.10 $\pm$ 0.02	119.4 $\pm$ 19.0
Drought	70.6 $\pm$ 3.5*	0.94 $\pm$ 0.14*	0.19 $\pm$ 0.05	0.09 $\pm$ 0.01	114.6 $\pm$ 29.9
Recovered	89.9 $\pm$ 4.8	0.51 $\pm$ 0.12	0.21 $\pm$ 0.07	0.09 $\pm$ 0.02	86.1 $\pm$ 17.8
<i>Peltandra virginica</i>					
Control	87.9 $\pm$ 1.0	0.03 $\pm$ 0.01	0.91 $\pm$ 0.21	1.05 $\pm$ 0.30	87.3 $\pm$ 3.6
Drought	84.3 $\pm$ 3.4	0.12 $\pm$ 0.03*	0.37 $\pm$ 0.07*	1.81 $\pm$ 0.32	90.0 $\pm$ 7.1
Recovered	84.1 $\pm$ 3.1	0.59 $\pm$ 0.04*	0.62 $\pm$ 0.12	1.04 $\pm$ 0.23	90.7 $\pm$ 7.1
<u>Dicots</u>					
<i>Justicia americana</i>					
Control	95.9 $\pm$ 0.5	0.32 $\pm$ 0.04	0.66 $\pm$ 0.06	1.17 $\pm$ 0.21	92.6 $\pm$ 7.28
Drought	74.8 $\pm$ 1.9*	0.87 $\pm$ 0.25*	0.34 $\pm$ 0.05*	1.01 $\pm$ 0.19	61.3 $\pm$ 22.0
Recovered	90.1 $\pm$ 5.1	1.12 $\pm$ 0.11*	0.16 $\pm$ 0.06*	0.32 $\pm$ 0.11*	74.9 $\pm$ 15.0
<i>Saururus cernuus</i>					
Control	96.9 $\pm$ 2.4	0.34 $\pm$ 0.05	0.24 $\pm$ 0.05	0.37 $\pm$ 0.12	102.6 $\pm$ 9.8
Drought	88.2 $\pm$ 1.1*	0.96 $\pm$ 0.16*	0.20 $\pm$ 0.03	0.14 $\pm$ 0.05	82.9 $\pm$ 23.5
Recovered	92.9 $\pm$ 2.0	0.89 $\pm$ 0.16*	0.21 $\pm$ 0.03	0.17 $\pm$ 0.04	83.9 $\pm$ 19.2

**Table 2.** Plant water status parameters derived from pressure-volume isotherms on control and drought-recovered plants. Parameters include water fraction at turgor loss point ( $\theta_{\text{tlp}}$ ; %), symplastic water fraction ( $\theta_{\text{sym}}$ ; %), osmotic potential at full saturation ( $-\psi_{\pi}^{\text{sat}}$ ; MPa), osmotic potential at the turgor loss point ( $-\psi_{\pi}^{\text{tlp}}$ ; MPa), and bulk modulus of elasticity ( $\varepsilon$ ; MPa). Statistical differences from the control are indicated by asterisks ( $p < 0.05$ ; one-way ANOVA). Data is presented as means  $\pm$  1 S.E. (n = 8).

Species (treatment)	$\theta_{\text{tlp}}$	$\theta_{\text{sym}}$	$-\psi_{\pi}^{\text{sat}}$	$-\psi_{\pi}^{\text{tlp}}$	$\varepsilon$
<u>Monocots</u>					
<i>Carex alata</i>					
Control	0.79 $\pm$ 0.01	0.54 $\pm$ 0.03	1.11 $\pm$ 0.06	1.26 $\pm$ 0.09	1.69 $\pm$ 0.36
Recovered	0.77 $\pm$ 0.04	0.50 $\pm$ 0.04	0.37 $\pm$ 0.03*	0.67 $\pm$ 0.09*	0.56 $\pm$ 0.06*
<i>Juncus effusus</i>					
Control	0.84 $\pm$ 0.03	0.52 $\pm$ 0.05	0.33 $\pm$ 0.03	0.52 $\pm$ 0.04	0.71 $\pm$ 0.10
Recovered	0.86 $\pm$ 0.03	0.61 $\pm$ 0.05	0.36 $\pm$ 0.05	0.54 $\pm$ 0.08	1.64 $\pm$ 0.26*
<i>Peltandra virginica</i>					
Control	0.92 $\pm$ 0.01	0.82 $\pm$ 0.01	0.52 $\pm$ 0.02	0.84 $\pm$ 0.02	4.26 $\pm$ 0.88
Recovered	0.85 $\pm$ 0.03*	0.61 $\pm$ 0.05*	0.86 $\pm$ 0.08*	1.38 $\pm$ 0.11*	1.95 $\pm$ 0.47*
<u>Dicots</u>					
<i>Justicia americana</i>					
Control	0.85 $\pm$ 0.01	0.32 $\pm$ 0.06	1.15 $\pm$ 0.08	1.43 $\pm$ 0.10	1.91 $\pm$ 0.24
Recovered	0.92 $\pm$ 0.02*	0.77 $\pm$ 0.04*	0.69 $\pm$ 0.03*	1.11 $\pm$ 0.06*	4.18 $\pm$ 0.62*
<i>Saururus cernuus</i>					
Control	0.90 $\pm$ 0.01	0.38 $\pm$ 0.05	1.38 $\pm$ 0.15	1.58 $\pm$ 0.17	3.60 $\pm$ 0.45
Recovered	0.93 $\pm$ 0.01*	0.71 $\pm$ 0.01*	0.76 $\pm$ 0.01*	1.03 $\pm$ 0.06*	8.49 $\pm$ 0.99*

## Significance

### **Part 1: Drought Susceptibility in Emergent Wetland Angiosperms: a Comparison of Water Deficit Growth in Five Herbaceous Perennials**

As our basic understanding of how different wetland macrophytes respond to changing soil water tables increases with future studies (including both controlled greenhouse experiments and field investigations) we should be able to develop more generalized patterns necessary for predicting plant-drought interactions. This basic understanding is fundamental if we are to develop better working models on how wetlands will respond to changing environmental conditions that lead to decreased water supply - including mechanical disturbances associated with human activities and projected decreases in water availability due to climate change.

### **Part 2: Drought Tolerance Versus Drought Avoidance: A Comparison of Plant-Water Relations in Herbaceous Wetland Plants Subjected to Water Withdrawal and Repletion**

In contrast, the former case of increased cell wall rigidity in drought stressed plants would allow for decreases in both turgor and water potential with only a small decrease in plant water content. Furthermore, a change in cell wall elasticity requires far less energy than the metabolically driven alternative of increasing organic compatible solutes (Lo Gullo et al. 1986). Consequently, Corcuera et al. (2002) suggested that plants growing in soils low in both water and nutrient content would favor the lower energy process of cell wall modification. While all species studied in this investigation can be found in organically rich soils typical of many freshwater wetland habitats, *J. americana*, *J. effusus*, and *S. cernuus* are also commonly found in nutrient poor clay soils typical of central Piedmont reservoirs of North Carolina (Touchette et al. 2000).

## References

### **Part 1: Drought Susceptibility in Emergent Wetland Angiosperms: a Comparison of Water Deficit Growth in Five Herbaceous Perennials**

Beadle, C.L. (1985). Plant Growth Analysis. In Coombs, J., Hall, D.O., Long, S.P., Scurlock, J.M.O. (eds.), *Techniques in Bioproductivity and Photosynthesis*. Pergamon Press Ltd., New York, NY, p. 20-25.

Bonilla-Warford, C.M., and Zedler, J.B. (2002). Potential for using native plant species in stormwater wetlands. *Environmental Management* 29: 385-394.

Breshears, D.D., Cobb, N.S., Rich, P.M., Price, K.P., Allen, C.D., Balice, R.G., Romme, W.H., Kastens, J.H., Floyd, M.L., Belnap, J., Anderson, J.J., Myers, O.B., Meyer, C.W. (2005). Regional vegetation die-off in response to global-change-type drought. *Proceedings of the National Academy of Science* 102: 15144-15148.

- Brinson, M.M., and Malvarez, A.I. (2002). Temperate freshwater wetlands: types, status, and threats. *Environmental Conservation* 29: 115-133.
- Bush, D.E., Loftus, W.F., and Bass, O.L. (1998). Long-term hydrologic effects on marsh plant community structure in the southern everglades. *Wetlands* 18: 230-241.
- Collins, B.S., and Battaglia, L.L. (2001). Hydrology effects on propagule bank expression and vegetation in six Carolina bays. *Community Ecology* 2: 21-33.
- Conner, W., and Day, J. (1992). Water level variability and litterfall productivity of forested freshwater wetlands in Louisiana. *American Midland Naturalist* 128: 237-245.
- Dawson, T.P., Berry, P.M., and Kampa, E. (2003). Climate change impacts on freshwater wetland habitats. *Journal for Nature Conservation* 11: 25-30.
- De Steven, D., and Toner, M. (2004). Vegetation of upper coastal plain wetlands: environmental templates and wetland dynamics within a landscape framework. *Wetlands* 24: 23-42.
- Dodd, C. (1994). The effects of drought on population structure, activity, and orientation of toads (*Bufo quercicus* and *B. terrestris*) at a temporary pond. *Ethology, Ecology, and Evolution* 6: 331-349.
- Ferris, R., Sabatti, M., Miglietta, F., Mills, R.F., and Taylor, G. (2001). Leaf area is stimulated in *Populus* by free air CO<sub>2</sub> enrichment (POPFACE), through increased cell expansion and production. *Plant, Cell and Environment* 24: 305-315.
- Gaines, K., Bryan, A., and Dixon, P. (2000). The effects of drought on foraging habitat selection of breeding Wood Storks in coastal Georgia. *Waterbirds* 23: 64-73.
- Galatowitsch, S.M., Whited, D.C., Lehtinen, R., Husveth, J., and Schik, K. (2000). The vegetation of wet meadows in relation to their land-use. *Environmental Monitoring and Assessment* 60: 121-144.
- Gopal, B., and Junk, W.J., (2000). Biodiversity in wetlands: an introduction, In Gopal, B., Junk, W.J., and Davis, J.A. (eds.), *Biodiversity in Wetlands: Assessment, Function and Conservation*. Vol. 1, pp. 1-10. Backhuys, Leiden, the Netherlands.
- Holland, C.C., Honea, J., Gwin, S.E., and Kentula, M.E. (1995). Wetland degradation and loss in the rapidly urbanizing area of Portland, Oregon. *Wetlands* 15: 336-345.
- Hunt, R. (1990). *Basic Growth Analysis: Plant Growth Analysis for Beginners*. Unwin Hyman Ltd. London, UK.

- Jacobs, J.M., Mergelsberg, S.L., Lopera, A.F., Myers, D.A. (2002). Evapotranspiration from a wet prairie wetland under drought conditions: Paynes Prairie Preserve, Florida, USA. *Wetlands* 22: 374-385.
- Johnson, W.C., Boettcher, S.E., Poiani, K.A., Guntenspergen, G. (2004). Influence of weather extremes on the water levels of glaciated prairie wetlands. *Wetlands* 24: 385-398.
- Katovich, E., Becker, R., Byron, J. (2003). Winter survival of late emerging purple loosestrife (*Lythrum salicaria*) seedlings. *Weed Science* 51: 565-568.
- Kennedy, M.P., Milne, J.M., and Murphy, K.J. (2003). Experimental growth responses to groundwater level variation and competition in five British wetland plant species. *Wetlands Ecology and Management* 11: 383-396.
- Kentula, M.E., Gwin, S.E., and Pierson, S.M. (2004). Tracking changes in wetlands with urbanization: sixteen years of experience in Portland, Oregon, USA. *Wetlands* 24: 734-743.
- Kercher, S.M., and Zedler, J.B. (2004). Flood tolerance in wetland angiosperms: a comparison of invasive and noninvasive species. *Aquatic Botany* 80: 89-102.
- Kirkman, L., and Sharitz, R. (1993). Growth in controlled water regimes of three grasses common in freshwater wetlands of the southeastern USA. *Aquatic Botany* 44: 345-359.
- Mangan, J.M., Overpeck, J.T., Webb, R.S., Wessman, C., Goetz, A.F.H. (2004). Response of Nebraska sand hills natural vegetation to drought, fire, grazing, and plant functional type shifts as simulated by the century model. *Climate Change* 63: 49-90.
- Mulhouse, J.M., Burbage, L.E., Sharitz, R.R. (2005). Seed bank-vegetation relationships in herbaceous Carolina bays: responses to climatic variability. *Wetlands* 25: 738-747.
- Olson, R.A. (2004). Components, processes, and design of created palustrine wetlands. In: McKinstry, M.C., Hubert, W.A., and Anderson, S.H. (eds), *Wetland and Riparian areas of the Intermountain West*. University of Texas Press, Austin, TX, pp 185-215.
- O'Neal, M.E., Landis, D.A., and Isaacs, R. (2002). An inexpensive, accurate method for measuring leaf area and defoliation through digital image analysis. *Journal of Economic Entomology* 95: 1190-1194.
- Pepin, S., Plamondon, A., and Britel, A. (2002). Water relations of black spruce trees on a peatland during wet and dry years. *Wetlands* 22: 225-233.
- Petranka, J, Murray, S., and Kennedy, C. (2003). Responses of amphibians to restoration of a southern Appalachian wetland: Perturbations confound post-restoration assessment. *Wetlands*, 23: 278-290.

Pezeshki, S.R., Anderson, P.H., and Shields, F.D. (1998). Effects of soil moisture regimes on growth and survival of black willow (*Salix nigra*) posts cuttings. *Wetlands* 18: 460-470.

Rea, N., and Ganf, G.G. (1994). Water depth changes and biomass allocation in two contrasting macrophytes. *Australian Journal of Marine and Freshwater Research* 45: 1459-1468.

Rejmankova, E., Rejmanek, M., Djohan, T., and Goldman, C. (1999). Resistance and resilience of subalpine wetlands with respect to prolonged drought. *Folia Geobotanica* 34: 175-188.

Rubio, G., and Lavado, R.S. (1999). Acquisition and allocation of resources in two waterlogging-tolerant grasses. *New Phytology* 143: 539-546.

Sala, A., and Nowak, R.S. (1997). Ecophysiological responses of three riparian graminoid to changes in the soil water table. *International Journal of Plant Science* 158: 835-843.

Smith, S., Devitt, D., Sala, A., Cleverly, J., and Busch, D. (1998). Water relations of riparian plants from warm desert regions. *Wetlands* 18: 687-696.

Sommer, B., and Horwitz, P. (2001). Water quality and macroinvertebrate response to acidification following intensified summer droughts in a Western Australian wetland. *Marine and Freshwater Research* 52: 1015-1021.

Sorenson, L.G., Goldberg, R., Root, T.L., and Anderson, M.G. (1998) Potential effects of global warming on waterfowl populations breeding in the Northern Great Plains. *Climatic Change* 40: 343-369.

Stuedler, S., and Touchette, B. (2003). The effects of drought on the productivity and growth of the aquatic macrophyte, American water willow (*Justicia americana* L.). *Southeastern Biology* 50 (2): 121.

Van der Valk, A.G. (1981). Succession in wetlands: a Gleasonian approach. *Ecology* 62: 688-696.

Williams, L.L., and Hudak, P.F. (2005). Hydrology and plant survival in excavated depressions near an earthen dam in north-central Texas. *Environmental Geology* 48: 795-805.

## **Part 2: Drought Tolerance Versus Drought Avoidance: A Comparison of Plant-Water Relations in Herbaceous Wetland Plants Subjected to Water Withdrawal and Repletion**

- Amlin, N.M., and Rood, S.B. (2002). Comparative tolerances of riparian willows and cottonwoods to water table decline. *Wetlands* 22: 338-346.
- Ayoub, A., Khalil, M., and Grace, J. (1992). Acclimation to drought in *Acer pseudoplatanus* L. (sycamore) seedlings. *Journal of Experimental Botany* 43: 1591-1602.
- Blake, T.J., Bevilacqua, E., Zwiazek, J.J. (1991). Effects of repeated stress on turgor pressure and cell elasticity changes in black spruce seedlings. *Canadian Journal of Forest Research* 21: 1329-1333.
- Bray, E.A. (1993). Molecular responses to water deficits. *Plant Physiology* 103: 1035-1040.
- Brock, M.T., and Galen, C. (2005). Drought tolerance in the alpine dandelion, *Taraxacum ceratophorum* (Asteraceae), its exotic congener *T. officinale*, and interspecific hybrids under natural and experimental conditions. *American Journal of Botany* 92: 1311-1321.
- Clifford, S.C., Arndt, S.K., Corlett, J.E., Joshi, S., Sankhla, N., Popp, M., and Jones, H.G. (1998). The role of solute accumulation, osmotic adjustment and changes in cell wall elasticity in drought tolerance in *Ziziphus mauritiana* (Lamk.). *Journal of Experimental Botany* 49: 967-977.
- Coruera, L., Camarero, J.J., and Gil-Pelegrín, E. (2002). Functional groups in *Quercus* species derived from the analysis of pressure-volume curves. *Trees* 16: 465-472.
- Cowardin, L.M., Carter, V., Golet, F.C., and LaRoe, E.T. (1979). *Classification of Wetlands and Deepwater Habitats of the United States*. U.S. Government Printing Office, Washington, D.C. 103 pp.
- Davies, F.S., and Lakso, A.N. (1979). Diurnal and seasonal changes in leaf water potential components and elastic properties in response to water stress in apple trees. *Physiologia Plantarum* 46: 109-114.
- Dichio, B., Xiloyannis, C., Angelopoulos, K., Nuzzo, V., Bufo, S.A., and Celano, G. (2003). Drought-induced variations of water relations parameters in *Olea europaea*. *Plant and Soil* 257: 381-389.
- Gaiser, T., de Barros, I., Lange, F.M, Williams, J.R. (2004). Water use efficiency of a maize/cowpea intercrop on a highly acidic tropical soil as affected by liming and fertilizer application. *Plant and Soil* 263: 165-171.
- Gao, X., Zou, C., Wang, L., and Zhang, F. (2004). Silicon improves water use efficiency in maize plants. *Journal of Plant Nutrition* 27: 1457-1470.

Chen, X.M., Begonia, G.B., Alm, D.M., and Hesketh, J.D. (1993). Responses of soybean leaf photosynthesis to CO<sub>2</sub> and drought. *Photosynthetica* 29: 447-454.

Girma F.S., and Krieg, D.R., (1992). Osmotic adjustment in Sorghum. I. Mechanisms of diurnal osmotic potential changes. *Plant Physiology* 99: 577-582.

Grammatikopoulos, G. (1999). Mechanism of drought tolerance in two Mediterranean seasonal dimorphic shrubs. *Aust. J. Plant Physiol.* 26:587-593.

Holbrook, N.M, and Putz, F.E., (1996). From epiphyte to tree: Differences in leaf structure and leaf water relation associated with the transition in growth form in eight species of hemiepiphytes. *Plant Cell and Environment* 19: 631-642.

Huang, X.M., Huang, H.B., and Gao, F.F. (2000). The growth potential generated in citrus fruit under water stress and its relevant mechanism. *Scientia Horticulturae* 83: 227-240.

Joly, R.J. (1985). Techniques for determining seedling water status and their effectiveness in assessing stress. In Duryea, M.L. (ed.), *Evaluating Seedling Quality: Principles, Procedures, and Predictive Abilities of Major Tests*. Forest Research Laboratory, Oregon State University, Corvallis, OR. pp. 17-28.

Jones H.G., and Corlett, J.E. (1992). Current topics in drought physiology. *Journal of Agricultural Science* 119: 291-296.

Karamanos, A.J. (1984). Way of detecting adaptive responses of cultivated plants to drought: an agronomic approach. In Margaris, N.S., Arianoutsou-Farragitakiand, M., and Oechel, W.C. (eds.), *Being Alive on Land: Tasks for Vegetation Science*. W. Junk Pup., Hague, pp. 91-101.

Koide, R.T., Robichaux, R.H., Morse, S.R., and Smith, C.M. (1989). Plant water status, hydraulic resistance and capacitance. In Pearcy, R.W., Ehleringer, J., Mooney, H.A., and Rundel, P.W. (eds.), *Plant Physiological Ecology: Field Methods and Instrumentation*. Chapman and Hall, New York, pp. 161-178.

Kozlowski, T.T., Kramer, P.J., and Pallardy, S.G. (1990). *The Physiological Ecology of Woody Plants*. Academic Press, New York, USA

Kramer, P.J. (1983). *Water Relations of Plants*. Academic Pres, Inc. New York, NY.

Kramer, P.J., and Boyer, J.S. (1995). *Water Relations of Plants and Soils*. Academic Press, New York, USA.

Liu, F., Andersen, M.N., Jacobsen, S-E., Jensen, C.R. (2005). Stomatal control and water use efficiency of soybean (*Glycine max* L. Merr.) during progressive soil drying. *Environmental and Experimental Botany* 54: 33-40.

Lo Gullo, M.A., Salleo, S., and Rosso, R. (1986). Drought avoidance strategy in *Ceratonia siliqua* L. a mesomorphic-leaved tree in the xeric Mediterranean area. *Annals of Botany* 58: 745-756.

Marron, N., Delay, D., Petit, J.M., Dreyer, E., Kahlem, G., Delmotte, F.M., and Brignolas, F., (2002). Physiological traits of two *Populus x euramericana* clones, Luisa Avanzo and Dorskamp, during a water stress and re-watering cycle. *Tree Physiology* 22: 849-858.

Meinzer, F.C., Grantz, D.A., Goldstein, G., and Saliendra, N.Z. (1990). Leaf water relations and maintenance of gas exchange in coffee cultivars grown in drying soil. *Plant Physiology* 94: 1781-1787.

Mitch, W.J., and Gosselink, (1986). *Wetlands*. Ban Norstran Reinhold Co., New York, NY.

Mitch, W.J., Dorge, C.L., and Wiemhoff, J.R. (1979). Ecosystem dynamics and a phosphorus budget of an alluvial cypress swamp in southern Illinois. *Ecology* 60:1116-24.

Morgan, J.M. (1984). Osmoregulation and water stress in higher plants. *Annual Reviews in Plant Physiology* 35: 299-319.

Munns, R. (1988). Why measure osmotic adjustment? *Australian Journal of Plant Physiology* 15: 717-726.

Pavlik, B.M. (1984). Seasonal changes of osmotic pressure, symplasmic water content and tissue elasticity in the blades of dune grasses growing in situ along the coast of Oregon. *Plant, Cell and Environment* 7: 531-539.

Peltier, J.P., and Marigo, G. (1999). Drought adaptation in *Fraxinus excelsior* L.: Physiological basis of the elastic adjustment. *Journal of Plant Physiology*: 154: 529-535.

Reichstein, M., Tenhunen, J.D., Roupsard, O., Ourcival, J.M., Rambal, S., Miglietta, F., Peressoti, A., Pecchiari, M., Tirone, G., Valentini, R. (2002). Severe drought effects on ecosystem CO<sub>2</sub> and H<sub>2</sub>O fluxes at three Mediterranean evergreen sites, revision of current hypotheses? *Global Change Biology* 8: 999-1017.

Saito, T., and Terashima, I. (2004). Reversible decreases in the bulk elastic modulus of mature leaves of deciduous *Quercus* species subjected to two drought treatments. *Plant, Cell and Environment* 27: 863-875.

Savé, R., Peñuelas, J., Marfá, O., Serrano, L. (1993). Changes in leaf osmotic and elastic properties and canopy structure of strawberries under mild water stress. *Horticultural Science* 28: 925-927.

- Schulte, P.J. (1992). The units of currency for plant water status. *Plant, Cell and Environment* 15: 7-10.
- Schultz, H.R., and Matthews, M.A. (1993). Xylem development and hydraulic conductance in sun and shade shoots of grapevine (*Vitis vinifera* L.): evidence that low light uncouples water transport capacity from leaf area. *Planta* 190: 393-406.
- Serrano, L., and Peñuelas, J. (2005). Contribution of physiological and morphological adjustments to drought resistance in two Mediterranean tree species. *Biologia Plantarum* 49: 551-559.
- Slavík, B. (1974). *Methods of Studying Plant Water Relations*. Springer-Verlag, Berlin.
- Stuedler, S.E., and Touchette, B.W. (2003). The effects of drought on the productivity and growth of the aquatic macrophyte, American water willow (*Justicia americana* L.). *Southeastern Biology* 50 (2): 121.
- Touchette, B.W., Burkholder, J., and Glasgow, H. (2000). *American Water Willow (Justicia americana L.) and other Aquatic Vegetation in the Narrows Reservoir: Habitat Value and Responses to Human Influences*. NCSU Technical Report, Alcoa Power Generating Inc., Yadkin, NC.
- Tschaplinski, T.J., Tuskan, G.A., Gebre, G.M., Todd, D.E. (1998). Drought resistance of two hybrid *Populus* clones grown in a large-scale plantation. *Tree Physiology* 18: 653-658.
- Turner, N.C., Spurway, R.A., and Schulze, E.D. (1984). Comparison of water potentials measured by in-situ psychrometry and pressure chamber in morphologically different species. *Plant Physiology* 74: 316-319.
- Tyree M.T., and Hammel H.T. (1972). The measurement of the turgor pressure and the water relations of plants by the pressure-bomb technique. *Journal of Experimental Botany* 23: 267-282.
- White, D.A., Turner, N.C., and Galbraith, J.H. (2000). Leaf water relations and stomatal behavior of four allopatric *Eucalyptus* species planted in mediterranean southwestern Australia. *Tree Physiology* 20: 1157-1165.
- White, R.H., Engelke, M.C., Anderson, S.J., Ruemmele, B.A., Marcum, K.B., and Taylor, G.R. (2001). Zoysiagrass water relations. *Crop Science* 41: 133-138.
- Wilcox, D.A., Meeker, J.E., Hudson, P.L., Armitage, B.J., Black, M.G., Uzarski, D.G. (2002). Hydrologic variability and the application of index of biotic integrity metrics to wetlands: A great lakes evaluation. *Wetlands* 22: 588-615.

Wilson, J.R, Ludlow, M.M., Fisher, M.J., and Schulze, E.D. (1980). Adaptation to water stress of the leaf water relations of four tropical forage species. *Australian Journal of Plant Physiology* 7: 207-220.

Yin, C., Wang, X., Duan, B., Luo, J., Li, C. (2005). Early growth, dry matter allocation and water use efficiency of two sympatric *Populus* species as affected by water stress. *Environmental and Experimental Botany* 53: 315-322.

Zlatev, Z.S. (2005). Effects of water stress on leaf water relations of young bean plants. *Journal of Central European Agriculture* 6: 5-14.